

Research Article

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Biofertilizer Impacts on Cassava (*Manihot Esculenta Crantz*) Cultivation: Improved Soil Health and Quality, Igbariam, Nigeria

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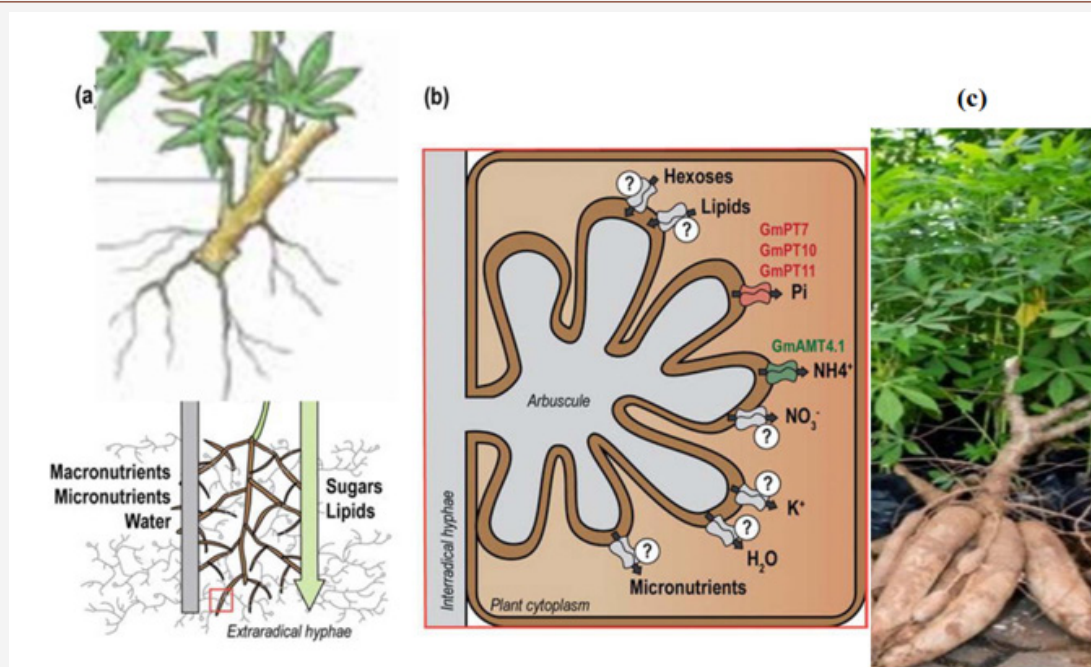
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Abstract

Cassava crop cultivation microbiome impacts by microbial biofertilizer is an integral function of the agro-ecology, its evolution (yield, soil health and quality) for sustainable agriculture. Increased yield and nutrient bioavailability powered by co-inoculation of microbial consortia adaptability to the soil ecology. Biofertilizer solves the traceability problem of chemical farm inputs, suitability and nutrient use efficiency as an integral function of the rhizosphere microbiome 'tailor' integrated soil fertility management that improves micronutrient and macronutrient via plant microbe interactions for soil health quality and crop degradation management. *Mycorrhizal spp* mutualistic symbiosis with plant roots satisfies the crop nutrients requirements. *Aspergillus spp*, *Bacillus spp* and *Clostridium spp* in the biofertilizer enhances potassium solubilization, a major challenge in cassava crop nutrition development of the biomass and mineralization. Soil organic matter improvement as a function of biofertilizer application help management of carbon sequestration and climate vulnerabilities.



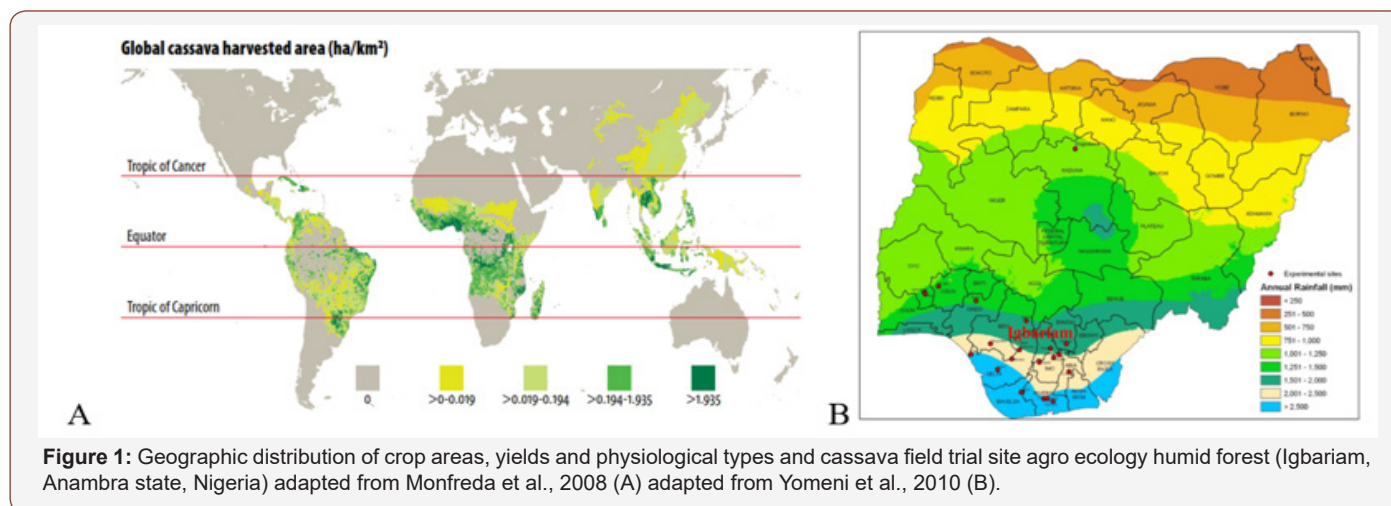
Keywords: Biofertilizer; Plant microbe interactions; Sustainable Food systems; Soil fertility; Climate change; Crop nutrient efficiency; Rhizosphere; Soil health and quality; Cassava (*manihot esculenta crantz*)

Introduction

Cassava cultivation constraints

Many diseases are caused by pathogens, whose damage symptoms appear on the leaves, stems and storage roots [1] during cassava cultivation. The common diseases of cassava are cassava mosaic disease, cassava bacterial blight, cassava anthracnose disease, cassava bud necrosis and root rot. Some of these diseases attack the leaves and stems of cassava plants while others attack the storage roots [2]. Cassava mosaic disease is caused by the African cassava mosaic virus which occurs inside the leaves and stems and causes yield reductions of up to 90 percent [3]. Economical damage

by diseases, pests and weeds of cassava is relatively moderate, although white flies can be a menace in some regions, if the problem is not identified early, and remedial action not implemented in a timely manner (Figure 1). Correct identification of the pest and an understanding of its behaviour, including its most vulnerable stages would provide insights into its management affects crops yield and development. Care must be then taken if pesticide application is contemplated, since there is the likelihood of high residual levels remaining in the product after harvest if an inappropriate formulation is not used.



Biopesticides can exert fungicidal, insecticidal, or nematocidal action via the microbial inoculate in the biofertilizer, a combination of them and possibly other auxiliary functions such as bird and mammal repellents or herbicides. According to recent classifications [4,5]. Bio-control action is due to multiple synergic mechanisms, generally including: i) production of antibiotics and other secondary metabolites (e.g., phenazines by *Pseudomonas spp.*, lipopeptides by *Bacillus spp.*, and hydrocyanic acid by Rhizobia); and ii) secretion of lytic and defense enzymes (e.g., chitinases, glucanases, peroxidases, polyphenol oxidases, and phenylalanine

ammonia lyases produced by *Trichoderma*, *Fusarium*, *Rhizoctonia*, *Serratia*, *Streptomyces* and *Bacillus* strains) [6,7]. The drawback of using living microorganisms is that their efficacy is often unpredictable under changing field conditions, and their fitness is reduced by the presence of an indigenous microbiota difficult to displace by non-native microorganisms [7,8]. Additionally, the antagonistic interactions occurring in formulations containing more than one microbial species limit their potential in integrated pest management strategies [9,10].

Biofertilizer formulation

Table 1: Categorization of general goals for agro-ecosystems.

Goal Type	General Goal	Key Controlling Variables
Economic Viability	High productivity	Genetic potential, weather, soil, management, economics
	Low cost of production	Yield potential*, input requirements*, input costs
	Low production risk	Market variation, production variation*
Stewardship	Preservation of productive land	Soil, climate, management
	Healthy animals	Feed quantity and quality*, disease
	High quality food and fiber	Chemical or microbial contamination*, composition*
Social	Viable local communities	Population size, economic viability, economic diversification
	Viable industry, institutions, and infrastructure	Profitability, size and resilience of industry
Environment	Clean water	Climate, soil, management
	Clean air	Climate, soil, management
	Wildlife habitat	Climate, soil, management

*Variables also influenced by soil proper.

A key advantage of beneficial microorganisms is to assimilate phosphorus for their own requirement, which in turn available as its soluble form in sufficient quantities in soil. *Pseudomonas*, *Bacillus*, *Micrococcus*, *Flavobacterium*, *Fusarium*, *Sclerotium*, *Aspergillus* and *Penicillium* have been reported to be active in the solubilisation process [11]. A phosphate-solubilizing bacterial strain *Micrococcus* sp. has polyvalent properties including phosphate solubilization and siderophore production [12]. Similarly, two fungi *Aspergillus fumigatus* and *A. Niger* were isolated from decaying cassava peels were found to convert cassava wastes by the semi-solid fermentation technique to phosphate biofertilizers [13]. *Aspergillus*, *Bacillus* and *Clostridium* are found to be efficient in potassium solubilization in

the soil and mobilize in different crops [14]. *Mycorrhizal* mutualistic symbiosis with plant roots (Figure 2) satisfies the plant nutrients demand [15] which leads to enhance plant growth and development and protect plants from pathogens attack and environmental stress [16]. *Pseudomonas aeruginosa* has been shown to withstand biotic and abiotic stresses [17]. Paul & Nair [18] found that *P. fluorescens* MSP-393 produces osmolytes and salt-stress induced proteins that overcome the negative effects of salt. Microbial inoculants genera in the OBD-Biofertilizer are isolated using the growth media in Table 1 from different agro biowaste and inoculated into the composted biofertilizer.

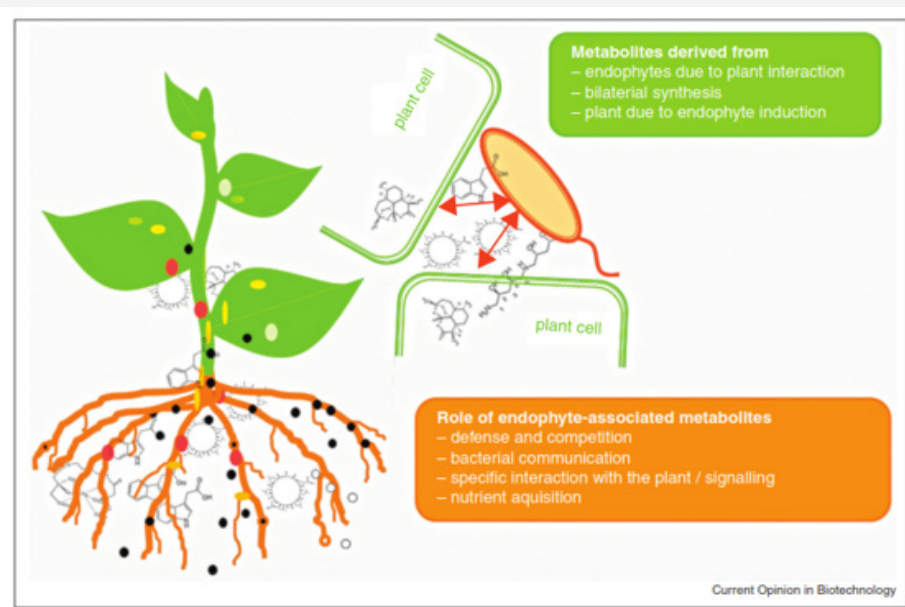


Figure 2: Schematic overview showing the different types of plant-endophyte interactions leading to the synthesis of metabolites, which are in many cases not produced by the macro- or micro symbiont alone or in different quantities, adapted from Sessitsch et al., 2014.

Climate change and soil biological health

It is commonly observed that applying only N or N + P can lead to a decline in particulate organic matter (>53 m fraction) and soil biological activity (soil respiration, microbial biomass C and N). These however improved significantly by moving towards balanced application through the addition of NPK or NPK+ organics [19]. Also, actual field studies on microbial diversity and activity are few. Contrary to a hypothesis that leaf litter produced under elevated CO₂ and having a high C: N ratio would be difficult to decompose, the microorganisms were found to adapt to changing soil carbon input under elevated CO₂ and there was no effect on their turnover and behaviour [20]. Expectedly, under 15 elevated CO₂, increased immobilization of fertilizer N by stimulation of mineralization (SMB) of soil organic matter (SOM) nitrogen was observed [21].

Thus, greater microbial demand for N (>27%) was observed under elevated CO₂ [22]. As warmer temperatures are maintained, the less efficient use of carbon by the microbes causes them to decrease in number, eventually resulting in less carbon dioxide being emitted into the atmosphere [23] via an agricultural soil vis-à-vis a desert soil (warmed in real world over time) attests

this reality. Mycorrhizal and N₂-fixing relationships are generally enhanced by CO₂ enrichment, but effects of warming are highly variable [24]. There are reports proving that soil resistance and resilience is linked to soil biodiversity [25] and 'higher' soil diversity protects the soil against ecosystem malfunctions under stress or disturbance: an 'insurance hypothesis' linked to soil biodiversity [26].

Unfortunately, some African soils lack essential nutrients. In Uganda, Kenya and Tanzania low yield of crops was attributed mainly to poor soil fertility [27]. For instance, Zn is deficient in most West African soils, especially the lowland areas [28] while plant viable P is unavailable in the iron-rich tropical soils of Africa due to low pH and high level of iron and aluminum oxides [29]. The soil lacks Ca, Mg and K, and when acidic, has a high level of free Mn, which is toxic to crops. Buhmann, et al. [30], some South African soils are deficient in K and P, making it unsuitable for cultivation. Africa has lower fertilizer consumption when compared to other regions of the world. In 2002, sub-Saharan Africa had about 8 kg/ha of fertilizer consumption which increased to 12 kg/ha in 2010 and 18 kg/ha in 2013 (Sommer et al., 2013). This is far below that of other regions of the world such as North America, South Asia, and

East Asia and Pacific which were estimated at 127.9 kg/ha, 151.8 and 337.0 kg/ha respectively (World Bank Fertiliser Consumption, 2013).

Sub-Saharan Africa fertilizer market lacks basic infrastructure for sustainability, efficient pricing and competition (Sommer et al., 2013). Biofertilizers should not be misunderstood for organic fertilizers such as compost, animal manure and plant manure or extracts [31,32]. However, whether the beneficial microbes improve crop accessibility to nutrients [6,33] or replenish soil nutrients (Shridhar, 2012; Thamer et al., 2011), if the overall nutrient condition of crop and soil has been improved, such substances containing the beneficial microorganisms are considered as biofertilizers [32]. The objectives are:

- How biofertilizer functional architecture links system design (microbial inoculant) impacts on the cassava crops nutrient use efficiency.
- To use the outcome indicators (crop yield, soil organic matter) as a determinant of soil health and quality and soil nutrient facility management.
- How the microbial inoculant impacts on the integrated soil management?
- What are the indicators of soil quality?

Indicator

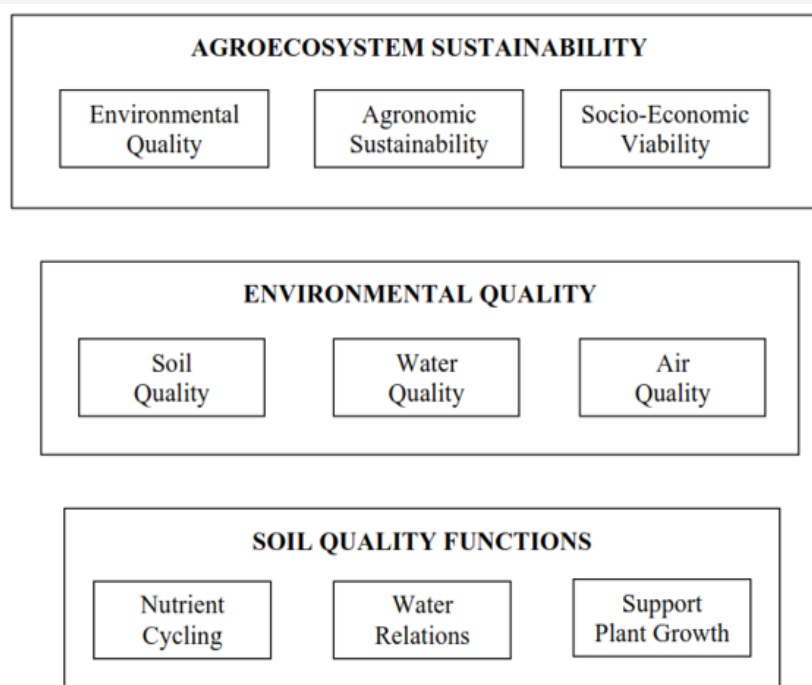


Figure 3: Nested hierarchy of agro-ecosystem sustainability showing the relationship of soil quality to the larger agro-ecosystem adapted from [64].

Biofertilizer is dependent variable is the variable being tested and measured in the cassava (independent variable or manipulated variable) field experiment. The independent variable (cassava crop) effect on the dependent variable is observed and recorded. Indicators can be used to communicate information on

Methodology

Biofertilizer functional models - soil health and quality

The environment-centric view (biofertilizer impacts) considers function as its effects (biofertilizer). The device-centric view considers function in term of internal parameters of the object (cassava crop physiology). The device-centric functions are the outcome (yield, soil health and quality) of the deployment of the environment centric functions. Eppinger and Browning, 2012 define. Underrating the biofertilizer system architecture of cassava crop cultivation within the agro-ecology, their relationships to crop development, evolution and outcome (yield, soil health and quality). Models are representations of the current understanding of a phenomenon or process of interest [34,35]. Functional models describe the relationship among variables using the simplest description of causal relations possible that still provides a useful description of the process or phenomenon [36]. A functional model would describe the components of the biofertilizer system and how they interact soils and crops cultivation. A mechanistic model would describe the properties of the biofertilizer contained in the components of the soil systems during cultivation. Information is also required on the driving forces that impact the variables controlling outcomes This driving force-outcome-response framework (or pressure-state- response framework) is widely used in environmental assessment [37].

indicators are often slow to respond but are directly related to the issue and are useful for assessment and planning. Response indicators communicate information on the extent to which remedial actions are implemented. Response indicators respond quickly, but their effects are not evident until much later. Indicators may communicate information on level, change or structure [38]. An indicator of structure provides information on industry or policy structures related to driving force (e.g., average farm size) or response (e.g., proportion of farms with an environmental farm plan). Water quality: watersheds with the greatest risk of non-point pollution are identified based on leaching and runoff vulnerability indices calculated for pesticides and nutrients (Figure 3).

For example, vulnerability indices for nutrients are obtained from estimates of excess nutrient levels (manure or commercial fertilizer sources) combined with estimates of leaching (based on precipitation and hydrologic factors) or estimates of run-off, Figure 2 reported by Kellogg et al. [39]. In the United States to develop soil ratings based on measured soil properties for the comparison of land management systems [40] and the approach, soil quality is considered an inherent property of the soil that can be determined from measurable soil attributes [41]. When a soil quality parameter declines below an acceptable limit, an appropriate response is required to increase soil quality. Acceptable limits depend on land use, soil characteristics, landform and climatic conditions. Many potential parameters of soil quality, measurable at various scales of assessment, have been proposed (Table 1). Wander & Bollero [42] concluded that particulate organic matter, mean wet weight diameter of aggregates, bulk density and penetration resistance may be good indicators of soil quality because they are sensitive to management and environmentally relevant.

Acton & Gregorich [43] defined soil quality as “the soil’s fitness to support crop growth without resulting in soil degradation or otherwise harming the environment”. Larson & Pierce [41] stated that “soil quality describes how effectively soils: 1) accept, hold, and release nutrients and other chemical constituents; 2) accept, hold, and release water to plants, streams and groundwater; 3) promote and sustain root growth; 4) maintain suitable biotic habitat; and 5) respond to management and resist degradation”. Karlen et al. [44] defined soil quality as “the capacity of a specific kind of soil to function, within natural or managed ecosystem boundaries, to sustain plant and animal productivity, maintain or enhance water and air quality, and support human health and habitation”.

Soil quality and health

Soil quality can be defined as the fitness of a specific kind of soil, to function within its capacity and within natural or managed ecosystem boundaries, to sustain plant and animal productivity, maintain or enhance water and air quality, and support human health and habitation [45]. Soil quality is related to soil functions and soil health concepts views soil as a finite and dynamic living resource [46]. Plant health is clearly a component of soil health but necessarily not of soil quality [47]. Baker & Cook [48] described the soils in which disease severity or incidence remains low, in spite of

the presence of a pathogen, a susceptible host plant and climatic conditions favorable for disease development, as suppressive soils. Soil biota like *arbuscular mycorrhizal* fungi play a significant role in improving plant nutrition but also act as bioprotectants against pathogens and toxic substances [49]. Thus, there is a considerable degree of overlap in the meaning of soil quality and soil health (Doran, 2002), though soil health perceptions tend to focus more on biotic components of soil [50]. Soil degradation or deterioration in soil health or quality implies loss of the vital functions of soil: (i) providing physical support, water and essential nutrients required for growth of terrestrial plants; (ii) regulation of the flow of water in the environment and (iii) elimination of the harmful effects of contaminants by means of physical, chemical and biological processes, i.e., environmental buffer or filter [38,51]. The quality and health of soil determine agricultural sustainability and environmental quality, which jointly determine plant, animal and human health [21,52].

Results and Discussion

Biofertilizer - mechanism of action

The absence of a population of degrading microorganisms can be overcome by the inoculation of the plant rhizosphere with pollutant degrading strains and biosurfactants during crop cultivation via biofertilizer. This approach successful in reducing the levels of benzene, ethylene, toluene xylenes, hydrocarbons, polychlorinated biphenyls and pesticides in polluted environments [50,53] especially in Africa poor soil profile. The rhizosphere is defined as the volume of the soil over which roots have influence, and which is shared with soil bacteria. Plants release exudates in the rhizosphere likely to serve as carbon source for microbes [54]. Consequently, rhizosphere microbes can promote plant health by stimulating root growth via production of plant growth regulators, enhance mineral and water uptake. Some bacteria, especially *fluorescent pseudomonads*, produce siderophores that have very high affinities for iron as compared to fungal siderophores [55] and can sequester this limited resource from other microflora thereby preventing their growth [56].

Earlier reports have demonstrated the importance of *P. fluorescens siderophores* in disease suppression [57,58], Figure 4. However, many endophytic bacteria are facultative plant colonizers and have to compete well in the rhizosphere before entering the plant [59] and might be therefore equipped with a rich arsenal of metabolites involved in defense as well as in interaction with the plant. Many bacteria with the capacity of colonizing plants utilize the nutrient niche of root surfaces in the rhizosphere and most of them might even actively switch from root surface to endophytic lifestyles [59,60]. These bacteria comprise several well characterized species of *Bacillus* and *Pseudomonas* and a number of metabolites, particularly lipopeptides synthesized by non-ribosomal peptide synthetases, have been described to be important for rhizosphere bacteria for antibiosis and for inducing plant defense mechanisms (Figure 5). Biofertilizer characteristics (Table 2) and biosurfactants (Table 3) applied in the filed cassava

cultivation requires no chemical pesticide. This was as a result of might be cassava plant-associated lifestyle requires adaptation to several niches, in which different metabolites act as signals for

interaction (communication) with the plant and host specific plants nutrient and crop protection.

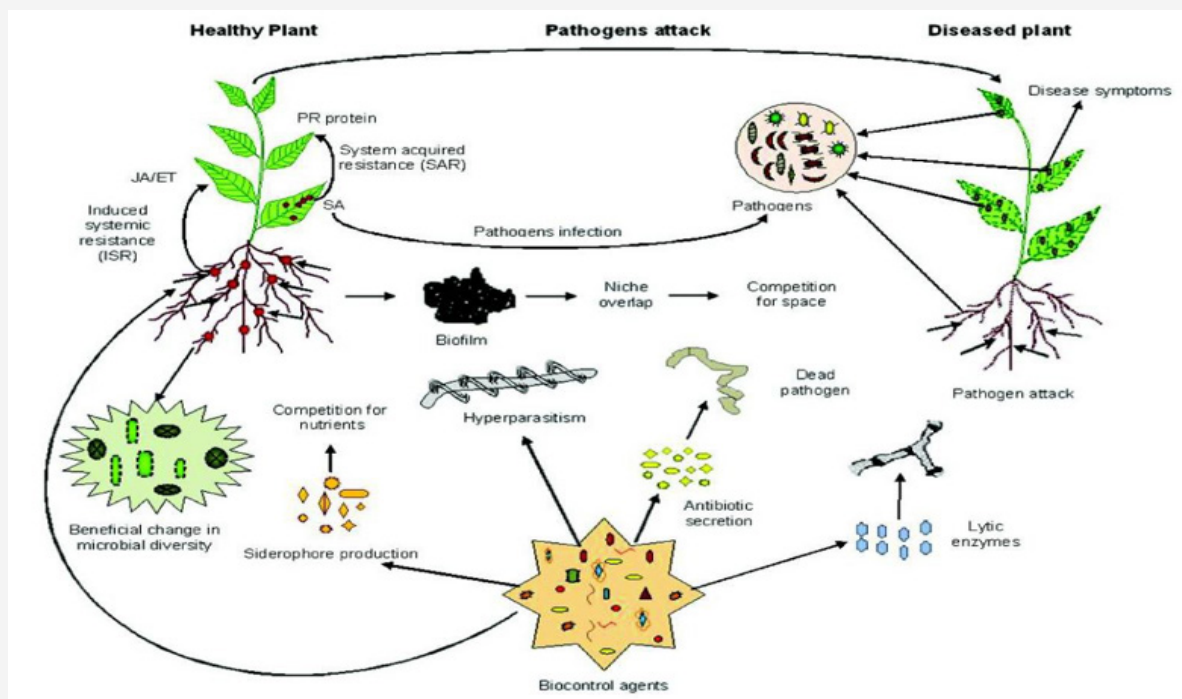


Figure 4: Mechanism of actions implemented by bio-control agents for management of plant diseases adapted from [98].

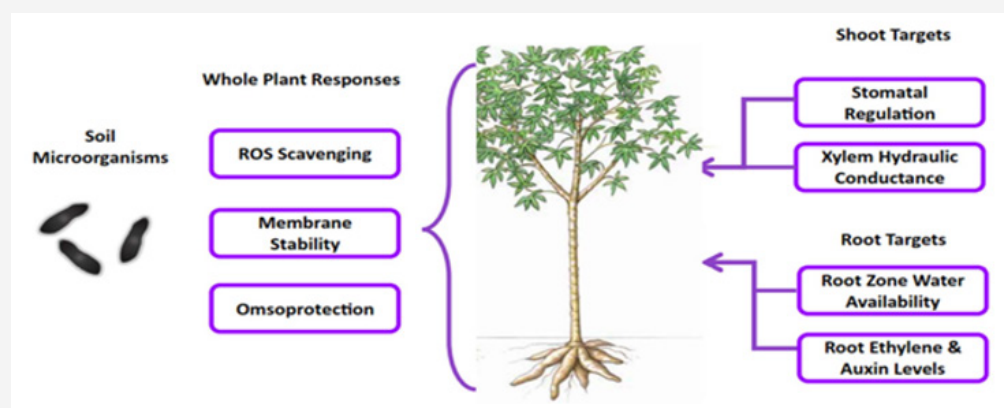


Figure 5: Summary of main key mechanisms targeted by microorganism-based bio stimulants in Table 3 and reactive oxygen species detoxification (ROS detox) enzymes might also ameliorate the plant-induced stress [138].

Table 2: Biofertilizer characteristic as integrated nutrient management during crop cultivation with references.

N/S	Characterization	Definition	Mechanisms	Crops Application	References
1	Biofertilizer	A substance which contains live microorganisms which, when applied on the seed, plant surface or the soil, colonizes the rhizosphere or the interior of the plant and promotes growth through increased supply or availability of primary nutrients for the host plant	a. Biological nitrogen fixation b. Utilization of insoluble forms of phosphorus	Maize, Soybean, Rice, Vegetables, Plantain, Horticulture	Vessey [32]; Somers, et al., [133] Fuentes-Ramírez & CaballeroMellado [104]
2	Phyto stimulator	Microorganism with the ability to produce or change the concentration of growth regulators such as indole acetic acid, gibberellic acid, cytokinin's and ethylene.	a. Production of phytohormones (auxins, cytokinin's and gibberellins). b. Decreased ethylene concentration (in the interior of the plant).		Lugtenberg, et al. [122]; Somers, et al. [133]

3	Biopesticide or biocontrol agent	Microorganisms that promote plant growth through the control of phytopathogenic agents, mainly for the production of antibiotics and antifungal metabolites.	a. Production of antibiotics (siderophores, HCN, antifungal metabolites)		Vessey [32]; Somers, et al. [133]; Chandler et al., 2008.
			b. Production of enzymes that degrade the		
			cellular wall of the fungi		
			c. Competitive exclusion		
			d. Acquired and Induced systemic resistance		
B	OBD-Biofertilizer	Ditto	ditto		ditto

Rhizobium and *Bacillus* were found to synthesize indole acetic acid (IAA) at different cultural conditions such as pH, temperature and in the presence of agro waste as substrate [61]. Ethylene, unlike other phyto-hormones, is responsible for the inhibition of growth of dicot plants [62]. It was found by Glick et al. [20] that PGPR could enhance the growth of plant by suppressing the expression of ethylene. Interestingly, a model was suggested in which it was shown that ethylene synthesis from 1-aminocyclopropane-1-carboxylate (ACC), an immediate precursor of ethylene, which is hydrolyzed by bacterial ACC- deaminase enzyme in the need of nitrogen and carbon source is also one of the mechanisms of induction of conditions suitable for growth, Figure 5. For plant-associated microorganisms introduced as bio-control agents into the rhizosphere or phyllosphere, the population of the microbial bio-control agent declines to background levels when the supporting plant dies, and it must be applied again with the next planting of that crop with the graphic narrative, Figure 5.

Increased yield and nutrient availability

The use of biofertilizers leads to separate accumulation of N, P and K in the soil, thereby maintaining soil nutrient balance [33,63]. Micronutrients (Zn, Fe, Mn, Cu and Mo) form insoluble complexes in the soil, which are not readily accessible by crops. Mahdi et al. [64], 75% of applied Zn forms insoluble complexes while plants use only about 1- 4% of total available Zn in the soil. However, *Rhizobium*, *Bradyrhizobium*, *Pseudomonas*, *Thiobacillus*, *Saccharomyces*, *Penicillium* and *Bacillus* can improve the uptake and availability of micronutrients in the soil [31,65]. In Fe immobilized soils, for example, bacteria siderophores solubilize and chelate Fe into complexes that can be easily absorbed by the plant roots [66]. *Trichoderma harzianum*, a fungal specie, can solubilize minerals such as metallic Zn and MnO by chelating and reducing mechanisms [67]. Vesicular Arbuscular Mycorrhiza (VAM) are also able to solubilize Zn, Fe, Mn, and Cu in agricultural soil [68,69]. This biofertilizer is important in areas experiencing high rate of potassium loss found in West Africa (Nigeria and Guinea Bissau) and East Africa (Burundi, Malawi, Kenya, Swaziland, Uganda and Rwanda) [70]. Another technique used by most plants to arrest P limiting situation is the plant-fungi symbiotic relationship (*ectomycorrhiza* and *endomycorrhiza*).

Plants develop increased root growth by the extension of the existing root systems through mycorrhizal association or hormonal stimulation (phyto-stimulation) effect [24]. Here, fungi hyphae are able to mobilize and make P available to the plants [6]. The

arbuscular mycorrhizal fungi (AMF) increase its exploitation of soil nutrients through specialized structures known as vesicles and arbuscules [71]. In addition, P is mobilized through the changes in sorption balance of soil solution caused by microbial biomass turnover in the rhizosphere. This leads to an increased mobility and uptake of organic P or orthophosphate ions [72]. Though some estimates on critical levels of soil organic matter (SOC) are available (e.g., Greenland et al. (1975)) considered 2% of SOC as the minimum requirement for maintenance of satisfactory soil aggregate stability and above which no further increases in productivity are achieved [21], the quantitative basis for such thresholds is limited (Loveland and Webb, 2003). Prasad et al. (2003), with particular reference to the Indian agriculture, considered soils with organic carbon (%) values < 0.5 as low fertility soils, 0.5 to 0.75 as medium fertility soils and >0.75 as high fertility soils. Magdoff (1998) reported potential crop yield increases of 12% for every 1% of soil organic matter based on his studies in USA. Soil quality indices are decision tools that effectively combine a variety of information for multi-objective decision making [73].

Enzymes as indicators of organic matter quality and microbial activity

Soil enzyme assays generally provide a measure of the potential activity, i.e., that encoded in the genotype, but this will rarely be ever expressed. Further, there are at least 500 enzymes and one has decided as to which enzymes would be the best indicators for soil quality [74], Table 4. Three enzymes viz., chitinase, phenol oxidase and phosphomonoesterase, as a group reflect relative importance of bacterial and fungi, as well as the nature of organic matter complex [75]. Phenol oxidase is produced primarily by white rot fungi, and is specific for highly recalcitrant organic matter, such as lignin [76]. Chitinase is a bacterial enzyme which converts chitin, a substance intermediate in its resistance to microbial metabolism produced by fungi and arthropods, into carbohydrates and inorganic nitrogen [77]. Phosphomonoesterase (acid phosphatase) activity is often correlated with microbial biomass [78,79], fungal hyphal length [58] and nitrogen mineralization [80]. Soil quality indicators would be useful to farmers and planners only if we know their critical limits, i.e., the desirable range of values of a given indicator that must be maintained for normal functioning of the soil.

Microorganisms affecting stress tolerance

Bacteria with the potential to act as bio stimulants (Table 3) have been isolated from a number of ecosystems with saline, alkaline,

acidic, and arid soils. These bacteria belong to several genera such as *Rhizobium*, *Bradyrhizobium*, *Azotobacter*, *Azospirillum*, *Pseudomonas* and *Bacillus* (Tables 3 & 4 respectively). Members of these genera have developed strategies to adapt and thrive under adverse conditions [81,82]. Amongst these adaptations, alterations to the composition of the cell wall and the ability to accumulate high concentrations of soluble solutes are common. These allow for

enhanced water retention and increased tolerance to osmotic and ionic stress. Cell wall composition is altered through enrichment for exopolysaccharides (EPS) and lipopolysaccharide-proteins and polysaccharide-lipids which form a protective biofilm on the root surface [83,84]. Plant growth-promoting rhizobacteria (PGPR) inoculated soils can ameliorate plant abiotic stress responses and narrated in detail in Figure 4.

Table 3: Microbial Metabolites Processes related to Plant Nutrient in Biofertilizer.

N/S	Genera	Microbe Species	Contribution	Disease Biocontrol	Crops	Metabolites	References
A	Bacteria	Agrobacterium	increased the NO ₃ and K uptake	Fusarium solan	Potato	antimicrobial metabolites like siderophores,	Idris, et al. [111]
		Azotobacter sp	consequently, the shoot and root	Botrytis cinerea	Beans, tomato	antibiotics, cyanides, fungal cell-wall-degrading enzymes	Lugtenberg & Kamilova [120]
		Bacillus sp	dry weights by 22 to 33 percent and	F. oxysporum		and gaseous products including ammonia.	Bertrand, et al. [92]
		Pseudomonas sp.	6 to 21 percent, respectively	Alternaria spp	Roots, leaves	Phenazines, pyrrolnitrin, pyoluteorin	Srivastava and Shalini, 2008
		Rhizobium sp		Sclerotium spp	rust on leaves, root rot and stem rot	and cyclic lipopeptides like viscosinamide.	
		Streptomyces sp		Colletotrichum lindemuthianum	Beans	Pseudobactin and pyoverdin.	Hillel [110]
		Enterobacter				Pyoverdine, pyochelin and its precursor salicylic acid chitinase and laminase.	
B.	Bacteria	Pseudomonus putida	Denitrification, methanogenesis, sulfidogenesis diseases and as therapeutic agents	Hydrocarbon Pollutants [Benzene, anthracene, hydrocarbons, PCBs]	biological remediation synoptic interaction of fermentative and acetogenic bacteria, with methnogens or	Oxygenase and peroxidases, pseudomicelle formation	Prescott, et al. [126]; Glazer & Nikaido, [107]; Kapley, et al. [115]
		Pseudomonas aerogenosa	Degrade hydrolysable tannins, diseases and as therapeutic agents	Agricultural/agro-industrial wastes	mineralization by amphipathic molecules	Plasmids, glycolipids, phospholipids, lipoproteins	Bhatta et al., 2012; Nitiema et al., 2010,
		Pseudomonas fluorescens		Antimicrobial activity	Decrease surface and interfacial tension	lipopeptides and polymeric compounds	Ray [130] Hamzah, et al. [108]
				Biofouling degradation	Decrease surface and interfacial tension	Reduction of interfacial tension	Chaillan, et al. [94]
				Antiviral activity		Rhamnolipid	Bhatia & Ichhpujani, 2005

		Bacillus sp	Bacillus cereus	Aromatics, long chain alkanes, phenol, cresol.	Sulphate reducers	Plasmids, glycolipids, phospholipids, lipoproteins	Cybulski, et al. [100]; Hamzah et al., 2013
		Azotobacter sp		Aromatics			Cybulski, et al. [100]
		Mycobacterium sp		Aromatics, branched hydrocarbons			Chatterjee, et al. [96]
				benzene, cycloparaffins			Jogdand [112]
		Streptomyces sp		phenoxyacetate, halogenated			Sunggyu [135]; Stanier et al., 1986
				hydrocarbon, diazinon			Jogdand [112], Kuiper, et al. [119]
		Streptococci sp	Degrade the most recalcitrant	Bioaccumulation of heavy metals	Mineralization	Beta-oxidation pathway	Rockne & Reddy [130], Schlegel, [135]
		Aspergillus species		Bioaccumulation of heavy metals	Mineralization	Beta-oxidation pathway	Gadd, [105]
							Gadd, [105]

Table 4: Key microbial metabolic processes related to plant nutrition.

Authors	Index Used/Proposed
Andrews, et al. [85]	Indices based on parameters related to entrance of water and plant growth
Bastida, et al. [38]	Microbiological index of soil degradation – dehydrogenase, water soluble carbohydrates, urease, water soluble carbon and respiration
Beck [86]	EAN – more enzyme activities (dehydrogenase, phosphatase, protease and amylase)
Dilly and Blume, 1990	As many as ten parameters
Doran and Parkin [102]	Index based on sustainable production, environmental quality and human and animal health
Doran and Parkin [102]	Soil quality index = function of (food and fibre production, erosivity, groundwater quality, surface water quality, air quality and food quality)
Kandeler and Eder [79]	Simple indices – quotients between enzymatic activity and microbial biomass
Kang, et al. [114]	Microbial index of soil (CHECK) based on microbial biomass C and N, potentially mineralizable N, soil respiration, bacterial population, mycorrhizal infection, and dehydrogenase and phosphatase activities
Karlen, et al. [73]	Soil quality index based on four soil functions: ability of soil to accommodate water entry, retain and supply water to plants, resist degradation and support plant growth
Klein and Paschke [117]	Total/active fungal and bacteria ratio – the ratio of total to active fungal plus bacterial biovolumes is divided by the ratio of the active fungal to bacterial biovolume
Parr, et al. [125]	Soil quality index based on different functions: soil properties, potential productivity, environmental factors, human and animal health, erodibility, biological diversity, food quality and safety and management inputs
Parr, et al. [125]	Soil quality index = function of (soil properties, potential productivity, environmental factors, human/animal health, erodibility, biological diversity, food quality/safety and management input
Puglisi, et al. [128]	Soil alteration index
Stefanic, et al. [88]	Biological index of soil fertility based on activity of two enzymes – dehydrogenase and catalase
Trasar-cepada et al., 1998;	Indices/equations based on parameters that reflect the total content of N or organic C
Harris, et al. [109]	Soil quality index based on three soil functions: ability to resist soil erosion, provide plant nutrients and provide a favorable root environment
Velasquez, et al. [139]	General indicator of soil quality based on abundance of 17 groups of macrofauna, eight soil chemical properties (extractable P, total P, exchangeable K, Mg, Ca, Na and pH, six physical properties (bulk density, real density, porosity, moisture content, shear strength, penetration resistance, soil morphological features and organic C fractions

Soil quality indices

Soil quality indices are decision tools that effectively combine a variety of information for multi-objective decision making [73].

A number of soil quality and fertility indices have been proposed [85-88], none identifies state of soil degradation that affects its functionality. Bastida et al. [38], building on the approach of Andrews et al. [85], suggested microbiological degradation index.

Scholars (Table 5) have appreciated and recommended the use of soil quality indices, reservations about their utility have also been expressed. Many a times the concepts associated with soil quality are used in close association with the concepts of sustainability, leading to a degree of confusion and inappropriate use of the term

soil quality [89] Table 6. Even though the importance of evaluation of soil quality is being increasingly realized, there is yet no global consensus on how this should be defined. While the notion of soil quality includes soil fertility, soil productivity, resource sustainability and environmental quality [90-115].

Table 5: Key microbial metabolic processes related to plant nutrition.

Element	Biochemical Process	Microbial Genes	Soil Enzymology Literature	Culture-Independent References	Culture-Dependent References
Nitrogen	Nitrogen fixation	nifD, nifH, nifK		Reganold et al., 2010; Xue et al., 2013	Bremer, et al. 1990
	Protein depolymerization	apr, npr, sub	Mader et al., 2002	Rasche et al., 2014	Kohler, et al. [118]
	Urea catabolism	ureA, ureB, ureC	Dick et al., 1988; Bowles et al., 2014	Reganold et al., 2010; Fierer et al., 2012, Xue et al., 2013	Kohler, et al. [118]
Phosphorous	Phosphate ester cleavage	phoA, phoD, phoX, ACPase, glpQ, ushA, appA, phyA, phyB	Mader et al., 2002; Garcia-Ruiz et al., 2008	Fraser et al., 2015	Kohler, et al. [118]
	Phosphonate breakdown	phnJ, phnX		Bergkemper et al., 2016	Schmalenberger, et al. [131]
Sulfur	Sulfate ester cleavage	asIA, asfA	Garcia-Ruiz et al., 2008	Schmalenberger et al [131]	
	Sulfonate breakdown	ssuD			Kertesz & Mirleau, 2004

Table 6: Ecological functions of soil (FAO, 1995) and their indicators.

Ecological Functions of Soil	Indicators of Proper Functioning
Production function	High levels of crop yields and incomes
Biotic environmental	High levels of species richness and functional dominance of
function/living space function	beneficial organisms – high levels of crop yields and incomes and high-quality food and habitation
Climate-regulative function/storage function	High levels of carbon stocks and slow rates of greenhouse gas emissions
Hydrologic function	Adequate availability of water/reduced risks floods
Waste and pollution control C-function	High levels of crop yields and incomes and high-quality food and habitation

Conclusion

In Nigeria and developing countries analysis of physical, chemical and biological characteristics of soil simultaneously is required to evaluate sustainability/ unsustainability of different management practices, most studies in developing countries have looked at physical and chemical characteristics only. Methodology of assessment of soil quality or soil health are still not in place for agro-ecology for crops microbiomes integrated soil nutrient management [116-132]. Biofertilizer accentuate the capacity of a soil to function by the improvement of the soil organic matter and designate cassava agro-ecology as 'suppressive' soils, Figure2 [133-140].

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Conflict of Interest

No conflict of interest.

References

- Miskito M, Braima J, Nnodu E, Legg J, Wydra K, et al. (2000) Disease Control in Cassava Farms. Wordsmithes Printers, Lagos, IITA, pp. 1-15.
- Olugbenga O. Ade Oluwa, Ofoso Budu, Brian Ssebunya (2011) Research Institute of Organic Agriculture, Switzerland.
- IITA (2008) Starting a Cassava Farm - IPM Field Guide for Extensions Agents. Technical Leaflet No.6 In: New Perspectives and Approaches in Plant Growth-Promoting Rhizobacteria Research. pp. 329-339.
- Czaja K, Góralczyk K, Strucinski P, Hernik A, Korcz W, et al. (2015) Biopesticides -towards increased consumer safety in the European Union. Pest Manag Sci 71(1): 3-6.
- Olson S (2015) An analysis of the biopesticide market now and where it is going. Outlook Pest Manag. Ortiz-Castro, Randy Hexon Angel Contreras-Cornejo, Lourdes 26(5): 203-206.
- Leahy J, Mendelsohn M, Kough J, Jones R, Berckes N (2014) Biopesticide oversight and registration at the U.S. environmental protection agency. In: Coats (Ed.), Biopesticides: State of the Art and Future Opportunities. ACS symposium series American Chemical Society, Washington, pp. 3-18.
- Parnell JJ, Berka R, Young HA, Sturino JM, Kang Y, et al. (2016) From the lab to the farm: an industrial perspective of plant beneficial micro-organisms. Front Plant Sci 7: 1110.
- Neeraja C, Anil K, Purushotham P, Suma K, Sarma P, et al. (2010a) Biotechnological approaches to develop bacterial chitinases as a bioshield against fungal diseases of plants. Crit Rev Biotechnol 30(3): 231-241.
- Gadhav KR, Hourston JE, Gange AC (2016) Developing soil microbial inoculants for pest management: can one have too much of a good thing? J Chem Ecol 42(4): 348-356.

10. Xu XM, Jefries P, Pautasso M, Jeger MJ (2011) Combined use of bio-control agents to manage plant diseases in theory and practice. *Phytopathology* 101(9): 1024-1031.
11. Pindi PK, Satyanarayana SDV (2012) Liquid microbial consortium- a potential tool for sustainable soil health. *J Biofertil Biopest* 13: 1-4.
12. Dastager SG, Deepa CK, Pandey A (2010) Isolation and characterization of novel plant growth promoting *Micrococcus* sp NII-0909 and its interaction with cowpea. *Plant Physiol Biochem* 48(12): 987-992.
13. Ogbo FC (2010) Conversion of cassava wastes for biofertilizer production using phosphate solubilizing fungi. *Bioresour Technol* 101(11): 4120-4124.
14. Mohammadi K, Yousef Sohrabi Y (2012) Bacterial Biofertilizers for sustainable crop production: A review. *J Agric Biol Sci* 7(5): 307-316.
15. Kogel KH, Franken P, Huckelhoven R (2006) Endophyte or parasite- what decides? *Curr Opin Plant Biol* 9(4): 358-363.
16. Lamabam PS, Gill SS, Tuteja N (2011) Unravelling the role of fungal symbionts in plant abiotic stress tolerance. *Plant Signal Behav* 6(2): 175-191.
17. Pandey PK, Yadav SK, Singh A, Sarma BK, Mishra A, et al. (2012) Cross-Species Alleviation of Biotic and Abiotic Stresses by the Endophyte *Pseudomonas aeruginosa* PW09. *J Phytopathol* 160(10): 532-539.
18. Paul D, Nair S (2008) Stress adaptations in a plant growth promoting Rhizobacterium (PGPR) with increasing salinity in the coastal agricultural soils. *J Basic Microbiol* 48(5): 1-7.
19. Manna MC, A Swarup, RH Wanjari, HN Ravankar, B Mishra, et al. (2005) Long term effect of fertilizer and manure application on soil organic carbon storage, soil quality and yield sustainability under sub-humid and semi-arid tropical India. *Field Crops Res* 93: 264-280.
20. Van Ginkel JH, Gorissen A (1998) In situ decomposition of grass roots as affected by elevated atmospheric carbon dioxide. *Soil Sci Soc Am J* 62: 951-958.
21. Gorissen A, Cotrufo MF (1999) Elevated carbon dioxide effects on nitrogen dynamics in grasses, with emphasis on rhizosphere processes. *Soil Sci Soc Am J* 63, 1695-Haberern J (1992) Viewpoint: a soil health index, *J Soil Water Conserv* 47(6).
22. Williams MA, Charles W Rice Clenton E Owensby (2001) Nitrogen competition in a tall grass prairie ecosystem exposed to elevated carbon dioxide. *Soil Sci Soc Am J* 65: 340-346.
23. Allison, S. D, Wallenstein MD, Bradford MA (2010) Soil-carbon response to warming dependent on microbial physiology. *Nature Geoscience* 3: 336-340.
24. Pritchard SG (2011) Soil organisms and global climate change. *Plant Pathology* 60(1): 82-99.
25. Garbeva P, Veen VJA, Elsas VJD (2004) Microbial diversity in soil: Selection of microbial populations by plant and soil type and implications for disease suppressiveness. *Annu Rev Phytopathol* 42: 243-270.
26. Yachi S, Loreau M (1999) Biodiversity and ecosystem productivity in a fluctuating environment: The insurance hypothesis. *Proc National Academy of Sciences* 96(4): 1463-1468.
27. Keino L, Baukya F, Ngetich W, Otinga AN, Okalebo JR, et al. (2015) Nutrients Limiting Soybean (glycine max I) growth in acrisols and ferralsols of Western Kenya. *PLoS ONE* 10(12): 1-20.
28. Abe SS, Buri MM, Issaka RN, Kiepe P, Takatsuki T (2010) Soil fertility potential for rice production in West African Lowlands. *Japan Agricultural Research Quarterly: JARQ* 44(4): 343-355.
29. De Valerla AW, Bake, A (2016) Micronutrient management for improving harvests, human nutrition, and the environment Scientific Project, Assigned by Food and Business Knowledge Platform. Wageningen: Wageningen University.
30. Buhmann C, Beukes D, Turner, D (2006) Plant nutrient status of soils of the Lusikisiki area, Eastern Cape Province. *South African Journal of Plant and Soil* 23(2): 93-98.
31. Ahsan ML, Ali A Ahmed I (2012) Biofertiliser a highly potent alternative to chemical fertilizers: Uses and prospects. *Journal of Chemical Engineering and Biological Science* 6(4): 10-23.
32. Vessey JK (2003) Plant growth promoting rhizobacteria as biofertilizers. *Plant Soil* 255(2): 571-586.
33. Egamberdiyeva D (2007) The effect of plant growth promoting bacteria on growth and nutrient uptake of maize in two different soils. *Applied Soil Ecology* 36(2-3): 184-189.
34. Addiscott TM (1993) Simulation modelling and soil behaviour. *Geoderma* 60(1): 15-40.
35. Yaalon DH (1994) On models, modeling, and process understanding. *Soil Science Society of America Journal* 58: 1276.
36. Dörner D (1996) The logic of failure. Metropolitan Books, New York, NY. 222 pp.
37. McRae T, Smith CAS, Gregorich LJ (2000) Environmental sustainability of Canadian agriculture: report of the agri-environmental indicator project. Agriculture and Agri-Food Canada, Ottawa, ON, pp. 1-232.
38. Bastida F, Moreno JL, Hernandez T, Garcia C (2006) Microbiological degradation index of soils in a smiarid climate. *Soil Biology and Biochemistry* 38(12): 3463-3473.
39. Kellogg RL, Wallace S, Alt K, Gos DW (1997) Potential priority watersheds for protection of water quality from nonpoint sources related to agriculture. 52nd Annual SWCS Conference. Toronto.
40. Karlen D L, Andrews S S, Doran J W (2001) Soil quality: Current concepts and applications. *Advances in Agronomy* 74: 1-40.
41. Larson WE, Pierce FJ (1994) The dynamics of soil quality as a measure of sustainable management. In J W Doran, DC Coleman, DF Bezdicek, BA Stewart, eds. *Defining soil quality for a sustainable environment*. Soil Science Society of America, Madison, WI, USA 35: 37 - 51.
42. Wander MM, Bollero G (1999) Soil quality assessment of tillage impacts in Illinois. *Soil Science Society of America Journal* 63(4): 961-971.
43. Acton DF, Gregorich LJ (1995) The health of our soils - towards sustainable agriculture in Canada. Centre for Land and Biological Resources Research, Research Branch, Agriculture and Agri-Food Canada, Ottawa, ON.
44. Karlen DL, Mausbach MJ, Doran JW, Cline RG, Harris RF, et al. (1997) Soil quality: a concept, definition and framework for evaluation. *Soil Science Society of America Journal* 61: 4-10.
45. Arshad MA, Martin, S (2002) Identifying critical limits for soil quality indicators in agroecosystems. *Agriculture, Ecosystems and Environment* 88(2): 153-160.
46. Doran JW, Zeiss MR (2000) Soil health and sustainability: managing the biotic component of soil quality. *Applied Soil Ecology* 15: 3-11.
47. Karlen D L, Mausbach MJ, Doran JW, Cline RG, Harris RF, et al. (1997) Soil quality: a concept, definition, and framework for evaluation. *Soil Science Society of America Journal* 61: 4-10.
48. Baker KF, Cook RJ (1974) Biological Control of Plant Pathogens. American Phytopathology Society, San Francisco, pp. 433.
49. Jeffries P, Gianinazzi S, Perotto S (2003) The contribution of arbuscular mycorrhizal fungi in sustainable maintenance of plant health and soil fertility. *Biol Fertil Soils* 37: 1-16.
50. Johnson DL, Maguire KL, Anderson DR, McGrath SP (2004) Enhanced dissipation of chrysene in planted soil: the impact of a rhizobia inoculum. *Soil Biology & Biochemistry* 36: 33-38.
51. Costanza R, Norton BG, Haskell BD (1992) *Ecosystem Health: New Goals for Environmental Management*. Island Press, Washington, DC.
52. Doran JW (2002) Soil health and global sustainability: translating science into practice. *Agriculture, Ecosystems and Environment* 88(2): 119-127.
53. Yousaf S, Ripka K, Reichenauer T, Andria V, Afzal M, et al. (2010a) Hydrocarbon degradation and plant colonization by selected bacterial

- strains isolated from Italian ryegrass and birds foot trefoil. *Journal of Applied Microbiology* 109(4): 1389-1401.
54. Olson PE, KF Reardon, EAH Pilon-Smits (2003) Ecology of rhizosphere bioremediation. In: S.C. McCutcheon and JL Schnoor (Ed.) *Phytoremediation: transformation and control of contaminants*. John Wiley and Sons, Inc, Hoboken NJ, pp. 317-353.
 55. O Sullivan DJ, O Gara F (1992) *Microbiol Rev* 56: 662-667.
 56. Kuc J (1995a) Phytoalexins, stress metabolism and disease resistance in plants. *Annu Rev Phytopathol* 33: 275-297.
 57. Costa JM, Loper JE (1994) *Mol Plant-Microbe Interact* 7: 440-448 pp.
 58. Leong SA, Expert D (1989) *Plant - Microbes Interactions, Molecular and Genetic Perspectives*, McGraw-Hill, New York 3: 62-83.
 59. Compant S, Clement C, Sessitsch A (2010) Plant growth-promoting bacteria in the rhizo- and endosphere of plants: their role, colonization, mechanisms involved and prospects for utilization. *Soil Biol Biochem* 42: 669-678.
 60. Rosenblueth M, Martinez-Romero E (2006) Bacterial endophytes and their interactions with hosts. *Mol Plant-Microbe Interact* 19: 827.
 61. Sudha M, Gowri RS, Prabhavati P, Astapriya P, Devi SY, et al. (2012) Production and optimization of indole-acetic-acid by indigenous micro flora using agro waste as substrate. *Pakistan J Biological Sci* 15(1): 39-43.
 62. Ansari MW, Trivedi DK, Sahoo RK, Gill SS, Tuteja N (2013) A critical review on fungi mediated plant responses with special emphasis to *Piriformospora indica* on improved production and protection of crops. *Plant Physiol Biochem* 70: 403-410.
 63. Adesemoye A, Torbert H, Kloepper J (2008) Enhanced plant nutrient use efficiency with PGPR and AMF in an integrated nutrient management system. *Can J Microbiol* 54(10): 876-886.
 64. Joylata laishram, KG Saxena RK, Maikhuri, KS Rao (2012) Soil Quality and Soil Health: A Review *International Journal of Ecology and Environmental Sciences* 38(1): 19-37.
 65. Adeleke R A, Cloete TE, Bertrand A, Khasa DP (2010) Mobilisation of potassium and phosphorus from iron ore by ectomycorrhizal fungi. *World Journal of Microbiology and Biotechnology* 26(10): 1901-1913.
 66. Mathew A, Eberl L, Earlier AL (2014) A novel siderophore-independent strategy of iron uptake in the genus *Burkholderia*. *Molecular Microbiology* 91(4): 805- 820.
 67. Altomare C, Norvell W, Bjorkman T, Harman G (1999) Solubilisation of phosphates and micronutrients by the plant-growth-promoting and biocontrol fungus *Trichoderma harzianum* Rifai 1295-22. *Appl Environ Microbiol* 65(7): 2926-2933.
 68. Martino E, Perotto S, Parsons R, Gadd GM (2003) Solubilization of insoluble inorganic zinc compounds by ericoid mycorrhizal fungi derived from heavy metal polluted sites. *Soil Biology and Biochemistry* 35(1): 133- 141.
 69. Pal S, Singh H, Farooqui A, Rakshit A (2015) Fungal biofertilisers in Indian agriculture: Perception, demand and promotion. *Journal of Eco-friendly Agriculture* 10(2): 101-113.
 70. Hartemink AAA, Lungu O, Naimi M, Okoth P, Smaling E, et al. (2012) African soils: Their productivity and profitability of fertilizer use. In: J Kihora, D Fatondji, JW Jones, G Hoogenboom, R Tabo, Bationo A (Eds.), *Improving soil fertility recommendation in Africa using decision support system for agro-technology transfer*, New York, pp. 19-42.
 71. Leigh J, Hodge A, Fitter AH (2009) Arbuscular mycorrhizal fungi can transfer substantial amounts of nitrogen to their host plant from organic material. *New Phytologist* 181(1): 199-207.
 72. Adeleke R, Nwangburuka C, Oboirien B. (2017) Origins, roles and fate of organic acids in soils: A review. *South African Journal of Botany* 108: 393-406.
 73. Karlen DL, Stott DE (1994) A framework for evaluating physical and chemical indicators of soil quality. In: Doran, J, Coleman DC, Bezdicsek DF, Stewart BA (Editors) *Defining Soil Quality for a Sustainable Environment*. Soil Science Society of America, Madison, WI 35: 53-72.
 74. Schlöter M, Dilly O, Munch JC (2003) Indicators for evaluating soil quality. *Agriculture, Ecosystems and Environment* 98(1-3): 255-262.
 75. Gai C, Boerner REJ (2007) Effects of ecological restoration on microbial activity, microbial functional diversity, and soil organic matter in mixed-oak forests of southern Ohio, USA. *Applied Soil Ecology* 35: 281-290.
 76. Carlisle MJ, Watkinson SC (1994) *The Fungi*. Academic Press, NY, pp. 482.
 77. Hanzlikova A, Jandera A (1993) Chitinase and changes of microbial community in soil. *Folia Microbiol.* 38(2): 159-160.
 78. Clarholm M (1993) Microbial biomass P, Labile P, and acid phosphatase activity in the humus layer of a spruce forest, after repeated additions of fertilizer. *Biology and Fertility of Soils* 8: 1281-1333.
 79. Kandeler E, Eder G (1993) Effect of cattle slurry in grassland on microbial biomass and on activities of various enzymes. *Biology and Fertility of Soils* 16: 249-254.
 80. Decker KLM, Boerner REJ, Morris SJ (1999) Scale-dependent patterns of soil enzyme activity in a forested landscape. *Canadian Journal of Forest Research* 29: 232-241.
 81. Selvakumar G, Joshi P, Mishra PK, Bisht JP, Gupta HS (2009) Mountain aspects influence the genetic clustering of psychrotolerant phosphate solubilizing *Pseudomonads* in the Uttarkhand Himalayas. *Curr Microbiol* 59(4): 432-8.
 82. Upadhyay SK, Singh DP, Saikia R (2009) Genetic diversity of plant growth promoting rhizobacteria from rhizospheric soil of wheat under saline conditions. *Curr Microbiol* 59(5): 489-96.
 83. Sandhya V, Ali AS, Grover M, Reddy G, Venkateswarlu B (2009) Alleviation of drought stress effects in sunflower seedlings by the exopolysaccharides producing *Pseudomonas putida* strain GAP-P45. *Biol Fertil Soils* 46(1): 17- 26.
 84. Zahran HH (1999) Rhizobium-legume symbiosis and nitrogen fixation under severe conditions and in an arid climate. *Microbiol Mol Biol Rev* 63(4): 968-89.
 85. Andrews SS, Karlen DL, Mitchell JP (2002) A comparison of soil quality indexing methods for vegetable production systems in Northern California. *Agriculture, Ecosystems and Environment* 90(1): 25-45.
 86. Bationo, Beck T (1984) Methods and application of soil microbiological analysis at the Landesanstalt für Bodenkultur und Pflanzenbau (LBB) in Munich for the determination of some aspects of soil fertility. In: Nemes MP, Kiss S, Papacostea P, Stefanic C, Rusan M (eds.), *Fifth Symposium on Soil Biology*, pp. 13-20.
 87. Karlen DL, Gardner JC, Rosek MJ (1998) A soil quality framework for evaluating the impact of CRP. *J prod Agric* 11: 56-60.
 88. Stefanic F, Ellade G, Chirnageanu J (1984) Researches concerning a biological index of soil fertility. In: Nemes MP, Kiss S, Papacostea P, Stefanic C, Rusan M (Eds.), *Fifth Symposium on Soil Biology*, pp. 35-45.
 89. Sojka RE, Upchurch DR (1999) reservations regarding the soil quality concept. *Soil Sci Soc Am J* 63(5): 1039-1054.
 90. Adeleke R, Cloete E, Khasa D (2010) Isolation and identification of iron ore-solubilising fungus. *South African Journal of Science* 106(9-10): 1-6.
 91. Bennet JW, Wunch KG, Faison B D (2002) Use of fungi biodegradation. *Manual of environmental microbiology 2002*, 2nd ed., ASM Press: Washington, DC, pp. 960-971.
 92. Bertrand TF, Fredric T, Robert N (2004) Production and partial characterization of a thermostable amylase from Ascomycetes yeast strain isolated from starchy soil. McGraw-Hill Inc, New York, pp. 53-55.
 93. Bezdicsek DF, Stewart BA (eds.), *Defining soil Quality for a Sustainable Environment*, SSSA Special Publication No.35, ASA and SSSA, Madison, WI, pp. 53-72.
 94. Chaillana F, Flècheb A, Burya E, Phantavongsa Y-hui, Saliot A, et al. (2004) Identification and biodegradation potential of tropical aerobic hydrocarbon-degrading microorganisms. *Res. Microb* 155(7): 587- 595.
 95. Chandrasekaran B, Josephson JR (2000) Function in device representation, *Engineering with Computers* 16: 162-177.

96. Chatterjee S, Haldar S, Asakura M, Yamasaki S, Balasubramanian T (2008) Molecular identification and phylogenetic status of marine bacillus associated with coral sediment, showing antibacterial effects against human pathogens. *Ann. Microbiol* 58(2): 309-312.
97. Chatterjee S, Mukherjee A, Agniswar S, Roy p (2012) Bioremediation of lead by lead resistant microorganisms isolated from industrial sample. *Advances in Bioscience and Biotechnology* 3: 280-285.
98. Cook RJ, Veseth RJ (1991) Wheat health management. APS Press, St. Paul, MN, pp. 152.
99. Cotter PD, Hill C, Ross RP (2005) Bacteriocins: developing innate immunity for food. *Nat Rev Microbiol* 3(10): 777-788.
100. Cybulski Z, Dzuirla E, Kaczorek E, Olszanowski A (2003) The influence of emulsifiers on hydrocarbon biodegradation by Pseudomonadacea and Bacillaceae strains. *Spill Science and Technology Bulletin* 8(5-6): pp 503-507.
101. Dilly O, Blume HP (1998) indicators to assess sustainable land use with reference to soil microbiology. *Advances in GeoEcology* 31: 29-39.
102. Doran JW, Parkin TB (1994) Defining and assesming soil quality. In: Doran JW, Coleman DC, Bezdieck DF, Stewart BA, (Eds.), *Defining Soil Quality for a sustainable environment*, Madison, WI, Soil Sci Soc Am 35: 3-21 (special publication).
103. Doran JW, Parkin TB (1996) Quantities indicators of soil quality: a minimum data set. In: Doran JW, Jones AJ (Eds.), *Methods for assessing Soil quality*, SSSA Special Publication NO.49, SSSA, Madison, WI, pp. 25-37.
104. Eppinger SD, Browning TR (2012) Design structure matrix methods and applications, MIT press, Fuentes-Ramírez LE, Caballero-Mellado J (2006) Bacterial biofertilizers. In: Z.A. Siddiqui (Ed). *PGPR: Biocontrol and Biofertilization*. Springer, Netherlands, pp. 143-172.
105. Gadd GM (1986) The uptake of heavy metals by fungi and yeasts: the chemistry and physiology of the process and applications for biotechnology, in *Immobilisation of Ions by Bio-sorption*, ed. By Eccles H and Hunt S. Ellis Horwood Ltd, Chichester, pp. 135-147.
106. Gil-Sotres F, Trasar-Cepeda C, Leiros MC, Seoane S (2005) different approaches to evaluating soil quality using biochemical properties. *Soil Biology and Biochemistry* 37(5): 877-887.
107. Glazer, AG Nikaldo H (2007) *Microbial Biotechnology Fundamentals of Applied Microbiology* 2nd Ed Gordon R (1994) *Bioremediation and its Application to Exxon Valdez Oil Spill in Alaska*.
108. Hamzah A, Rabu A, Azmy RFHR, Yusoff NA (2010) Isolation and characterization of bacteria degrading Sumandak and South Angsi oils. *Sains Malaysiana* 39(2): 161-168.
109. Harris RF, Karlen DL, Mulla DJ (1996). A conceptual framework for assesment and management of soil quality and health. In: Doran JW, Jones AL (Eds.), *Methods for assessing soil quality*. ASA and SSSA, Madison, WI 49: 61-82.
110. Hillel D, (2005) Thermal properties and processes. In: *Encyclopedia of Soils in the Environment*. D. Hillel JH, Hatfield DS, Powlson C, Rosenzweig KM, Scow MJ, Singer, DL Sparks Eds 4: 156-163.
111. Idris EES, Iglesias DJ, Talon M (2007) Tryptophan dependent production of indole-3-acetic 267 acid (IAA) affects level of plant growth promotion by *Bacillus amyloliquefaciens* FZB42. *Mol 268 Plant Microbe Interact* 20(6): 619-626.
112. Jogdand SN (1995) *Environmental biotechnology*, 1st Edition, Himalaya Publishing House, Bombay, India, pp. 104120.
113. Kandeler E, Gerber H (1988) Short-term assay of soil urease activity using colorimetric determination of ammonium. *Biology and Fertility of Soils* 6: 68-72.
114. Kang GS, Beri V, Sidhu BS, Rupela OP (2005) A new index to assess soil quality and sustainability of wheat-based cropping systems. *Biology and Fertility of Soils* 41: 389-398.
115. Kapley A, Purohit HJ, Chhatre S, Shanker R, Chakrabati T, et al. (1999) Osmotolerance and hydrocarbon degradation by a genetically engineered microbial consortium. *Biosour. Technol* 67(3): 241-245.
116. Karlen DL (1992) Soil and crop management effects on soil quality indicators. *American. J Alternative Agric* 7: 48-55.
117. Klein DA, Paschke MW (2000) A soil microbial community structural -functional index: the microscopy-based total/active/active fungal/ bacterial (TA/AFB) bio volumes ratio. *Applied Soil Ecology* 14(3): 257-268.
118. Kohler J, Caravaca F, Carrasco L, Roldan A (2007) Interactions between a plant growth-promoting rhizobacterium, an AM fungus and a phosphate-solubilising fungus in the rhizosphere of *Lactuca sativa*. *Appl Soil Ecol* 35: 480-487.
119. Kuiper I, Lagendijk EL, Bloemberg GV, Lugtenberg BJJ (2004) Rhizoremediation: a Beneficial Plant-Microbe Interaction. *Mol Plant Microbe Interact* 17(1): 6-15.
120. Lugtenberg B, Kamilova F (2009) Plant-Growth-Promoting Rhizobacteria. *Annual Review of Microbiology* 63: 541-556.
121. Lugtenberg B, Kamilova F (2009) Plant-Growth-Promoting Rhizobacteria. *Annual Review of Microbiology* 63: 541-556.
122. Lugtenberg BJJ, Chin A Woeng TFC, Bloemberg GV (2002) Microbe-plant interactions: principles and mechanisms. *Anton. Leeuw* 81: 373-383.
123. Mahdi SS, Hassan G, Samoan S, Rather H, Dar SA, et al. (2010) Biofertilizers in organic agriculture. *Journal Phytology* 2(10): 42-54.
124. Oldal B, Jevcsák I, Kecskés M (2002) A sziderofortermel o képesség szerepe *Pseudomonas*-törzsek növénypatogénantagonista hatásának biológiai vizsgálatában. *Biokémia* 26: 57-63.
125. Parr JF, Papendick RI, Hornick SB, Meyer RE (1992) Soil quality: attributes and relationship to alternative and sustainable agriculture. *Am. J. Alternative Agric* 7: 5-11.
126. Prescott LM, Harley JP, Klein DA (2002) *Microbiology. Fundamentals of applied Microbiology* 2: 1012-1014.
127. Prescott MI, Harle JD, Klein DA (2002) *Microbiology of Food*. 5th Ed. McGraw-Hill Ltd, New York, USA, pp. 964-976.
128. Puglisi E, Nicelli M, Capri E, Trevisan M, Attilio AM (2005) A soil alteration index based on phospholipids fatty acids. *Chmosphere* 61(11): 1548-1577.
129. Randlett DL et al (1996) Elevated atmospheric carbon dioxide and leaf litter chemistry: Influence on microbial respiration and net nitrogen mineralization. *Soil Sci Soc Am J* 60: 1571-1577.
130. Ray G (1994) *Bioremediation and its application to Exxon Valdez Oil Spill in Alaska*. Ray's Environmental Science Web Site Rockne, Karl and Reddy, Krishna (2003). *Bioremediation of Contaminated Sites*. University of Illinois at Chicago.
131. Schmalenberger A, Hodge S, Bryant A, Hawkesford MJ, Singh BK, Kertesz MA (2008) The role of variovorax and other comamonadaceae in sulphur transformations by microbial wheat rhizosphere communities exposed to different sulphur fertilization regimes. *Environ Microbiol* 10(6): 1486-1500.
132. Shimomura Y, M Yoshioka, H Takeda, Y Umeda, T Tomiyama (1998) Representation of design object based on the functional evolution process model, *Journal of Mechanical Design* 120(2): 221-229.
133. Somers E, Vanderleyden J, Srinivasan M (2004) Rhizosphere bacterial signalling: a love parade beneath our feet. *Crit Rev Microbiol* 30(4): 205-240.
134. Sparling GP, Schipper LA, Bettjeman W, Hill R (2004) Soil quality monitoring in New Zealand: practical lessons from a 6-year trial. *Agriculture, Ecosystems and Environment* 104(3): 523-534.
135. Sunggyu L (1995) Bioremediation of polycyclic aromatic hydrocarbon contaminated soil. *Journal of Cleaner Production* 3: 255.
136. Trasar Cepeda C, Leiros C, Gil Stotres F, Seoane S (1997) Towards a biochemical quality index for soils: an expression relating several biological and biochemical properties. *Biology and Fertility of Soils* 26: 100-106.

137. Van Oosten VR, Bodenhausen N, Reymond P, Van Pelt JA, Van Loon LC, et al. (2008) Differential effectiveness of microbial induced resistance against herbivorous insects in Arabidopsis. *Molecular Plant-Microbe Interactions* 21(7): 919-930.
138. Van Oosten MJ, Olimpia P Stefania De Pascale, Silvia S, Maggio A (2017) The role of biostimulants and bioeffectors as alleviators of abiotic stress n crop plants *Chem Biol Technol Agric* 4: 5.
139. Velásquez E, C Pelosi, D Brunet, M Grimaldi, M Martins, et al. (2007) This ped is my ped: visual separation and NIRS spectra allow determination of the origins of soil macro-aggregates. *Pedobiologia* 51(1): 75-87.
140. Wander MM, SJ Traina, BR Stinner and SE Peters (1994) Organic and conventional management effects on biologically active soil organic matter pools. *Soil Sci Soc Am J* 58: 1130-1139.