



Research Article

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Silver Nanoparticles Protect Complement C3 from Degradation by *Staphylococcus aureus* and *Serratia marcescens*

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Abstract

Bacterial pathogens utilize strategic mechanisms to circumvent or evade host immune defenses. Opportunistic pathogens like *Staphylococcus aureus*, *Serratia marcescens*, and *Klebsiella (Enterobacter) aerogenes*, can cause local and systemic infections. Evasion of host immune defenses can also lead to sepsis. The complement system plays a critical role in killing pathogens and signaling other host immune processes to activate. Blocking or impeding the function of critical complement proteins like C3 can lead to bacterial evasion of host defenses and dissemination through the system. Previous studies show that the function of proteases produced by *S. aureus* and *S. marcescens* can be inhibited by synthesized silver nanoparticles (AgNP). This study evaluated the ability of *S. aureus*, *S. marcescens*, or *K. aerogenes* to degrade human C3 and if nanoparticles could block the potential degradation of C3 by bacterial proteases. It is hypothesized that degradation of C3 by *S. aureus*, *S. marcescens*, and *K. aerogenes* could be prevented by AgNPs. To evaluate the hypothesis, bacteria were incubated with purified C3 in the presence and absence of AgNPs, followed by analysis through immunoblot analysis. Results show that *S. aureus* and *S. marcescens* degrade human C3, but *K. aerogenes* did not. Further analysis demonstrates that AgNP protects the integrity of C3 from degradation by *S. aureus* and partially protects C3 from *S. marcescens* degradation. Data from this study supports the hypothesis.

Introduction

Staphylococcus aureus is a Gram-positive bacterium commonly found as part of the normal flora but can be an opportunistic pathogen leading to nosocomial and community-acquired infections [1]. The emergence of multidrug resistant *S. aureus* called Methicillin-resistant *Staphylococcus aureus* (MRSA) is a concern for the medical and scientific communities. *S. aureus* produces hydrolytic enzymes and other virulence factors which degrade host tissues and evade host immune defense strategies [2-5]. *Serratia marcescens* is a Gram-negative opportunistic pathogen implicated in infections of immunocompromised individuals and is a leading cause of infections in burn victims [6,7]. *S. marcescens* is an organism

found in the respiratory and urinary tracts of hospitalized individuals and in the gastrointestinal tract of children. This pathogen produces virulence factors which too can cause destruction to host and evasion of the immune defenses [8-11]. *Klebsiella (Enterobacter) aerogenes* is a Gram-negative opportunistic pathogen associated with respiratory, urinary, and gastrointestinal infections [12]. All three opportunistic pathogens can cause very serious infections and complications in susceptible individuals.

Host immune defenses are critical for preventing microbial infections or dissemination of pathogens through the body. Some bacterial pathogens can destroy host tissues and disrupt normal

homeostatic and immune functions. The innate defense, including complement system, can become activated by various triggers including bacterial pathogens [13-15]. The complement components are proteins that may be subjected to bacterial degradation as a strategy of evading host defenses. If proper immune defenses are disrupted, pathogens can disseminate throughout the body leading to more serious infections and complications [13-15]. Previous studies show that silver nanoparticles can inhibit the protease activities of *S. aureus* and *S. marcescens* [16]. This study further examined if silver nanoparticles could protect complement C3 from degradation by *S. aureus*, *S. marcescens*, or *K. aerogenes*. Complement C3 protein is an important, critical component of the innate immune response which aids in phagocytosis of pathogens, opsonization, and complement cascade activation for clearance of pathogens. Upon activation, C3 is cleaved into C3a (anaphylatoxin) and C3b. This will then result in C5 cleavage leading to a powerful immune response [13-15]. In this study, the immunoprotective effect of nanoparticles on preventing C3 degradation was evaluated. Protecting C3 from degradation may help maintain proper immune defense against serious pathogens.

Material and Methods

Growth and preparation of bacteria

Bacteria *S. aureus*, *S. marcescens* and *K. aerogenes* were grown to stationary phase (optical density 600 (OD₆₀₀) of values over 1.0) in brain heart infusion broth (BHI) at 37° C for 20 hours. Bacterial cells were harvested by centrifugation and resuspended in IX Phosphate Buffered Saline (PBS) to an OD₆₀₀ of 0.6.

Synthesis of Silver Nanoparticles (AgNP)

Silver nanoparticles were synthesized similarly as previously described [16,17]. Briefly, silver nanoparticles (AgNP) were synthesized as follows. 0.01M AgNO₃ was placed in a round bottom flask and heated. 1% trisodium citrate was added and continued heating with stirring until the solution was an amber yellow color. The sample was removed from the heat immediately with continued stirring until it cooled to room temperature. A sample of the solution was evaluated in the UV-vis spec to confirm identity. Characteristic absorbance at 410nm was recorded and particles had an average diameter of 32nm as determined by Dynamic Light Scattering (DLS) spectrophotometry [16,17].

Treatment of Complement C3 protein with pathogens

Evaluation of bacterial impact on C3: To determine if pathogens can degrade complement C3 protein, 100µl of resuspended *S. aureus*, *S. marcescens* or *K. aerogenes* grown to stationary phase were incubated with 100 ng of commercially available C3 protein (TexGene) for a course of 6 hours at 37°C. After incubation, samples were centrifuged, and cell-free supernatants containing the C3 proteins were analyzed by immunoblot (Western blot) analysis. The negative control in this study was C3 incubated with 100µl of PBS

and no bacteria.

Evaluation of AgNP protection of C3 integrity: To evaluate if AgNP can protect C3 from bacterial degradation, 100µl of suspended bacteria (*S. aureus*, *S. marcescens* and *K. aerogenes*) were incubated with 100ng of C3 (TexGene) in the presence of 50 µl of silver nanoparticles for 6 hours at 37°C. Bacterial cells were removed via centrifugation and the cell-free fraction were separated by SDS-PAGE and then cleavage or degradation was evaluated by Western blot analysis. Controls for this study includes C3 incubated with Ag-NPs, and C3 incubated with PBS (no bacteria).

Western Blot Analysis: Experimental samples were prepared for SDS-PAGE and Western blot as follows: 65 µl of experimental sample, 10 µl of reducing agent (Life Technologies), 25 µl of LDS sample buffer (Life Technologies) then heated to 80oC for 10 minutes to denature samples. The sample proteins were separated using standard SDS-PAGE technique using a gradient 4-12% Bis-Tris gel (Life Technologies). After electrophoresis, the proteins were then transferred to PVDF membrane (Life Technologies) and C3 protein was probed using primary antibodies rabbit anti-C3 antibodies (TexGene). The Western Breeze Chromogenic Western blot kit (Life Technologies) was used to detect the integrity of the C3 protein. Instructions were followed according to manufacturer's recommendations (Life Technologies).

Results and Discussion

Bacterial degradation of complement C3

To evaluate if *S. aureus*, *S. marcescens*, and *K. aerogenes* could degrade C3, immunoblot (Western blot) analysis of C3 incubated with the three bacteria was done. The integrity of the intact 185-190kDa C3 bands was evaluated. Results show that in comparison to the untreated C3 control, *S. marcescens* completely degrades the complement C3 protein resulting in a smear with no observable bands around the 185-190 kDa (Figure 1). *S. aureus* incubation with C3 results in a reduction in the 185-190 kDa bands intensity compared to the untreated negative control (Figure 1). *K. aerogenes* did not appear to degrade C3 (Figure 1). These findings point to a possible strategy that is used by certain pathogens to escape immune defense. Normally one of the three complement pathways (classical, lectin, or alternative) are activated by a particular stimulus. No matter which complement pathway is triggered, they all converge at complement C3. Once activated, C5 becomes cleaved and activated which leads to the formation of the membrane attack complex. If complement C3 is degraded, the pathogen may escape elimination by the complement cascade which can lead to disease or damage by the pathogen. It would be beneficial for the host to preserve C3 integrity and function.

AgNP partially protects C3 from *S. marcescens*

C3 incubated with *S. marcescens* results in the loss of the C3 protein band integrity by the presence of visible smearing and complete degradation of C3 (Figure 2). C3 incubated with *S. marcescens*

in the presence of AgNP shows some intact 185-190kDa C3 complement protein bands. Degradation is not completely prevented, but some C3 is preserved (Figure 2). In contrast, *K. aerogenes* did not result in any C3 cleavage in the presence or absence of AgNP (Figure 3). This data suggest that AgNP can provide some protection of C3 from *S. marcescens*. Our lab is currently investigating if bime-

tallic gold-silver nanoparticles (AgAuNP) could provide better protection of C3. Previous studies from our lab show that AgAuNP can reduce the proteolytic activities of *S. marcescens* more than monometallic AgNP. If there is greater inhibition of proteolytic function, there may be less degradation of C3, preserving part of the host innate immune defense.

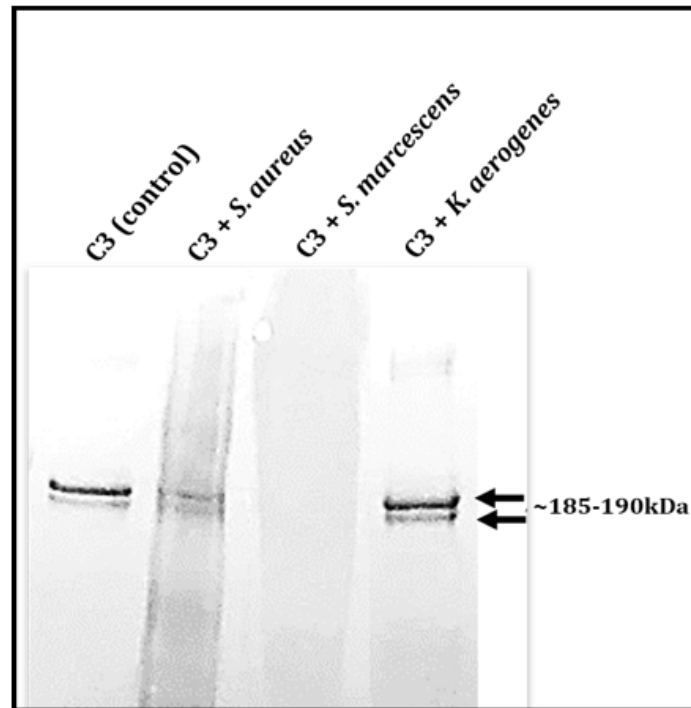


Figure 1: *S. aureus* and *S. marcescens* can degrade or cleave C3

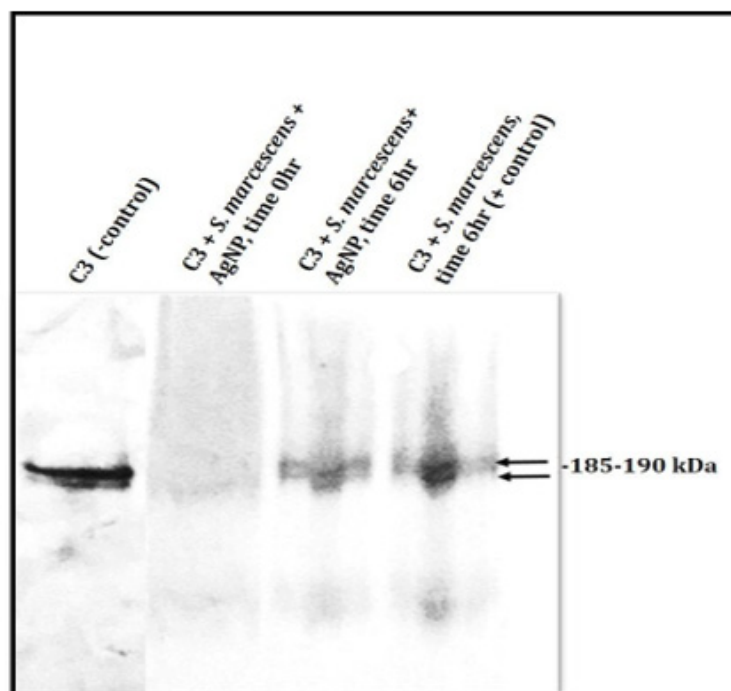


Figure 2: AgNP can protect C3 from *S. marcescens*-induced degradation.

AgNP protects C3 from *S. aureus*-induced degradation or cleavage

Results with *S. aureus* demonstrate that C3 is partially degraded or cleaved by *S. aureus* (Figure 4). After incubating C3 with *S. aureus* in the presence of AgNP, no visible detection of C3 degradation is observed in comparison to the untreated C3 control. The 185-190 kDa C3 bands appear to be preserved (Figure 4). Of special note is that without the presence of AgNP, *S. aureus* not only partially degrades or cleaves C3, but an immunoreactive band approximately 49kDa is also observed that is absent when AgNP is present (data

not shown). Studies in our lab are ongoing to investigate if C3 from total serum would be protected by AgNPs. These studies are currently evaluating C5 integrity, cleavage of C5 into C5a and C5b, and the downstream mechanisms involving the formation of the membrane attack complex and certain inflammatory reactions. We are also investigating the components of the specific components of the classical, alternative, or lectin pathways. Interestingly, other research has shown that *Staphylococcus pseudintermedius* was able to evade the host immune response by altering the function of complement and antibodies [18].

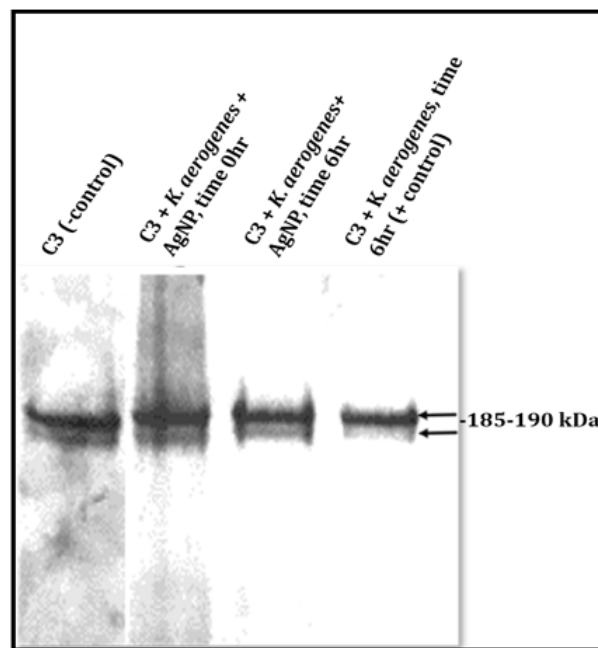


Figure 3: *K. aerogenes* does not degrade or cleave C3.

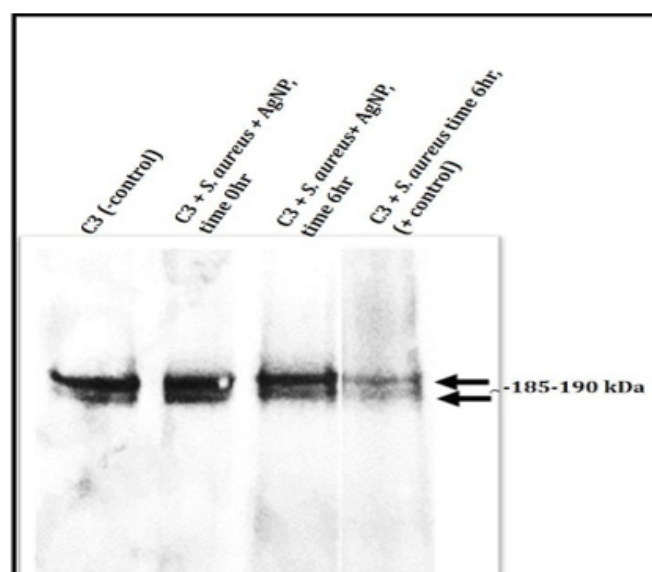


Figure 4: AgNP protects C3 from degradation by *S. aureus*.

Conclusion

Complement is a critical component of host innate immunity. Disruption in key complement components can have dire consequences including preventing the clearance of bacterial pathogens possibly leading to sepsis, and septic shock. Our findings show that AgNP can protect C3 from degradation and/or cleavage by *S. aureus* and *S. marcescens*. These mechanisms can also be investigated in other important human pathogens. In addition, the findings of this study can be used by the medical and scientific communities to identify strategies in combating complications caused by bacterial pathogens.

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Conflict of Interest

The authors have no conflict of interest.

References

- Lowy FD (1998) *Staphylococcus aureus* infections. N Engl J Med 339(8): 520-532.
- Pietrocola G, Nobile G, Rindi S, Speziale P (2017) *Staphylococcus aureus* manipulates innate immunity through own and host-expressed proteases. Front Cell Infect Microbiol 7:166.
- Tong SY, Davis JS, Eichenberger E, Holland TL, Fowler VG Jr (2015) *Staphylococcus aureus* infections: epidemiology, pathophysiology, clinical manifestations, and management. Clin Microbiol Rev 28(3): 603-661.
- Singh V, Phukan UJ (2019) Interaction of host and *Staphylococcus aureus* protease-system regulates virulence and pathogenicity. Med Microbiol Immunol 208(5): 585-607.
- Kantyka T, Shaw LN, Potempa J (2011) Papain-like proteases of *Staphylococcus aureus*. Adv Exp Med Biol 712: 1-14.
- Acar JF (1986) *Serratia marcescens* infections. Infect Control 7(5): 273-278.
- Hejazi A, Falkiner FR (1997) *Serratia marcescens*. J Med Microbiol 46(11): 903-912.
- Fredy J Romero, Luis A García, José A Salas, Mario Díaz, Luis M Quirós (2001) Production, purification and partial characterization of two extracellular proteases from *Serratia marcescens* grown in whey. Process Biochemistry 36(6): 507-515.
- Salamone PR, Wodzinski RJ (1997) Production, purification and characterization of a 50-kDa extracellular metalloprotease from *Serratia marcescens*. Appl Microbiol Biotechnol 48(3): 317-324.
- Shanks RM, Stella NA, Hunt KM, Brothers KM, Zhang L, et al. (2015) Identification of SlpB, a Cytotoxic Protease from *Serratia marcescens*. Infect Immun 83(7): 2907-2916.
- Zhang L, Morrison AJ, Thibodeau PH (2015) Interdomain contacts and the stability of serralyisin protease from *Serratia marcescens*. PloS one 10(9): e0138419.
- Wesevich A, Sutton G, Ruffin F, Park LP, Fouts DE, et al. (2020) Newly Named Klebsiella aerogenes (formerly *Enterobacter aerogenes*) is associated with poor clinical outcomes relative to other *Enterobacter* species in patients with bloodstream infection. J Clin Microbiol 58(9): e00582-620.
- Singer L, Colten HR, Wetsel RA (1994) Complement C3 deficiency: human, animal, and experimental models. Pathobiology 62(1): 14-28.
- Janssen BJ, Gros P (2007) Structural insights into the central complement component C3. Mol Immunol 44(1-3): 3-10.
- Doorduijn DJ, Bardoel BW, Heesterbeek DAC, Ruyken M, Benn G, et al. (2020) Bacterial killing by complement requires direct anchoring of membrane attack complex precursor C5b-7. PLoS Pathog 16(6): e1008606.
- Vanterpool J, Stuffle E, Vanterpool E (2021) Silver and gold-silver nanoparticles can serve as potential protease inhibitors for *Staphylococcus aureus* and *Serratia marcescens* hydrolytic enzymes. Sci J Biol & Life Sci 1(4): 1-4.
- Holden MS, Nick KE, Hall M, Milligan JR, Chen Q, et al. (2014) Synthesis and catalytic activity of pluronic stabilized silver-gold bimetallic nanoparticles. RSC Adv 4(94): 52279-52288.
- Sewid AH, Hassan MN, Ammar AM, Bemis DA, Kania SA (2019) *Staphylococcus pseudintermedius* Sbi paralogs inhibit complement and bind IgM, IgG Fc and Fab. PLoS One 14(7): e0219817.