



First Reported Case of Human Infection with *Chromobacterium Violaceum* in Gabon: Antibiotic Susceptibility Patterns and Treatment Outcome

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Abstract

Background: Mostly found in soil and still water in the subtropics and tropics, *Chromobacterium violaceum* is a Gram-negative bacillus that rarely infects humans. Nevertheless, when *Chromobacterium violaceum* infections do occur, they result in great distress because they are unrecognized and poorly addressed. The aim here is to describe what appears to be the first human case of *Chromobacterium violaceum* infection in Gabon.

Case presentation: A 12-month-old female was admitted to the paediatric ward of the Centre Regional Hospital Amissa Bongo of Franceville in Gabon with a fever that had been lingering for a week. The child was first put on antimalarial treatment for 3 days (following a positive thick blood smear for *Plasmodium falciparum*) combined with a 10-day prophylactic course of antibiotics (association of 500mg Ceftriaxone and Tobramycine).

Subsequently, a post-treatment cytobacteriological examination of urine (CBEU) was performed: a urine dipstick (UD) was carried out and then the urine was plated on Cystine-lactose-electrolyte-deficient (CLED) agar medium and incubated for 24 hours at 37°C.

Isolated bacterial colonies were identified with the Vitek 2 system (bioMérieux, France). Antibiotic susceptibility tests were run based on the Kirby-Bauer method (interpretation based on the breaking points given by the CLSI for *Escherichia coli*).

The UD revealed leukocyturia (30.103µL⁻¹) and greyish non-pigmented bacterial colonies (positive for catalase and oxidase) found on CLED (DGU=104UFC.mL⁻¹) were identified as *Chromobacterium violaceum*.

Antibiotic susceptibility tests exhibited resistance to beta-lactams (Ticarcillin, Ertapenem, Cefotaxime, Ceftazidime and Ceftriaxone), aminoglycosides (Gentamicin and Tobramycin), urinary quinolones (Nalidixic acid) and sulphonamides (Trimethoprim/Sulfamethoxazole).

Patient outcome: The infectious syndrome disappeared after the probabilistic treatment was adjusted post-CBEU and switched to Cefaclor 125 mg/5ml at a dosage of 25 mg/kg/day.

Conclusion: *Chromobacterium violaceum* seems to be a severe emerging human pathogen that requires prompt, adequate and lab results antibiotic-based treatment to prevent fatal outcomes.

Keywords: *Chromobacterium violaceum*; Urinary tract infection; Antibacterial resistance; Case report; Gabon

Introduction

Global climate change may lead to the rise of pernicious human infections caused by environmental bacteria, such as *Chromobacterium violaceum*, normally infecting animals. Among the *Chromobacterium* genus, *Chromobacterium violaceum* (*C. violaceum*) is the only species that is known to infect humans [1-3].

Over the years, this facultative anaerobic Gram-negative environmental proteobacterium has been responsible for over 200 cases of human infection worldwide, mostly in Southeast Asia, Australia, Africa, and Florida. Despite the scarcity of *C. violaceum* infections, they can lead to severe sepsis with a high risk of mortality particularly in immunocompromised individuals (G6PD deficit, diabetes, lymphogranuloma, etc.,) who appear to be more susceptible to this bacteremia [4,5]. In humans, *C. violaceum* infections can be challenging to treat due to antibacterial resistance.

Case Presentation

Patient's concerns and clinical findings: A 12-month-old female infant was admitted to the paediatric ward of the Centre Hospitalier Régional Amissa Bongo de Franceville (CHRA) for a prolonged fever lasting seven days. This patient had a history of hospitalization. He had been treated for malaria and an ENT infection (details of ENT infection not at disposal at the time this report was written) following episodes of febrile attacks at 2 and 4 months after birth. The patient had one week's history of fever before admittance to the paediatric ward. On admittance, the clinical examination was uneventful.

Table 1: Note: **AMC:** Amoxicillin/Clavulanic Acid; **KF:** Cefalotin; **FOX:** Cefoxitin; **CTX:** Cefotaxim; **ETP:** Ertapenem; **NA:** Nalidixic Acid; **CIP:** Ciprofloxacin; **PRL/TZP:** Piperacilin/Tazobactam; **OFX:** Ofloxacin; **TOB:** Tobramycin; **TI:** Ticarcilin; **IMI:** Imipenem; **AK:** Amikacin; **STX:** Trimethoprim/Sulfametoxazol; **CAZ:** Ceftazidim; **CN:** Gentamicin; **CRO:** Ceftriaxon; **S:** Susceptible; **R:** Resistant.

ATB	Disc charge (µg)	Interpretation
AMC	30	S
KF	30	S
FOX	30	S
CTX	5	R
ETP	10	R
NA	30	R
CIP	5	S
PRL/TZP	36	S
OFX	5	S

Primary diagnoses and interventions: The laboratory assessment highlighted the presence of an infectious syndrome: neutrophilia, a positive thick drop (parasitaemia not given here) and the C-reactive protein was 96 mg/L. The child has been put under injectable antimalarial treatment following national guidelines and probabilistic antibiotic therapy combining injectable ceftriaxone (C3G at a dose of 10 mg/kg/d for 10 days) with injectable Tobramycin (aminoglycoside at a dose of 3 mg/kg/d for 3 days) in direct intravenous. The clinical course of the malaria access was very satisfactory, but the infectious syndrome persisted. A CBEU carried out post-probabilistic antibiotic therapy came back positive and the child was switched to Cefaclor 125 mg/5 ml (oral C1G at a dose of 25 mg/kg per day).

The CBEU was performed as follows: a sample of urine was collected using a paediatric collection device under strict aseptic conditions. Following mixing, 10µL of urine was streaked onto a Cystine-lactose-electrolyte-deficient (CLED) agar medium and then, incubated at 37°C for 24 hours.

After 24 hours of incubation, whitish, smooth, shiny colonies appeared, catalase + oxidase +. The leucocyturia on the Kovaslide slide was around 30.103 leucocytes/µL. The urine germ count (UGC) was 104CFU/mL. Identification using the Vitek 2 automated system of bioMérieux (France) gave *C. violaceum* with 94% precision. Antibiotic susceptibility testing (AST) was performed using the Kirby-Bauer method also called the agar disc diffusion technique. The interpretation of AST was done according to CLSI recommendations and using the breaking points of *E. coli* (Table 1).

TOB	10	R
TI	75	R
IMI	10	S
AK	30	S
SXT	25	R
CAZ	10	R
CN	10	R
CRO	30	R

The results in Table 1 show that some antibiotics (AMC, KF, FOX, CIP, PRL/TZP, OFX, IMI and AK) could have been efficiently used against this strain of *C. violaceum*. The bacterium strain exhibited resistance to some antibiotics belonging to diverse families: TI, ETP, CTX, CAZ and CRO from the 3rd Generation Cephalosporins (C3G); CN and TOB from the Aminoglycosides; NA from the urinary Quinolones; and STX from the sulfonamides.

Outcome: The infectious syndrome disappeared after the probabilistic treatment was adjusted post-CBEU and switched to Cefaclor 125 mg/5ml at a dosage of 25 mg/kg/day. The child was then released and was doing great by the time we wrote this report.

Discussion

Only one case of *C. violaceum* was reported in the region of Central Africa and the Great Lakes so far [6], making the case here the second and the very first in Gabon. While most pigmented strains of *C. violaceum* have been described to be pathogenic to humans, some non-pigment strains can express the same pathogenic power [7-11].

The persistence of the infectious syndrome after clearing the malaria infection and especially after receiving probabilistic antibiotic therapy ("shot" of Ceftriaxone and Tobramycin) seems to underline to failure of the medication. The bacterium *C. violaceum* was identified following the CBEU and its antibiotic susceptibility tests showed that this bacterial strain was resistant to Ceftriaxone, which could further explain the persistence of the infectious syndrome despite the use of this compound by the patient. This was observed by another research group which also revealed that *C. violaceum* was as well resistant to Tobramycin [12]. However, more recent work has confirmed these findings by demonstrating that several commonly used antibiotics are inactive against this bacterial species, namely Penicillins and Cephalosporins, but the mechanisms that may underline this resistance need to be further explored. It would though appear that this resistance is of an acquired type and runs counter to the observations of several authors [7,13].

The resolution of the infectious syndrome following the lab CBEU report and the administration of a C1G as a treating compound suggested that this drug was indeed active on *C. violaceum*. Though our results confirmed bacterial strain sensitivity, it is still quite puzzling as it is unusual [14]. The most active antibiotics belong to the Carbapenem (Imipenem) and Fluoroquinolone (especially Ciprofloxacin) subfamilies [13,7, 15]. Responsiveness to one Carbapenem and two systemic Fluoroquinolones has been

frequently described by other authors [15] and of all the aminoglycosides tested; only Amikacin was found to be a good therapeutic choice to treat the infection, which is consistent with findings in the management of bacteremia due to this pathogen in Nepal [7].

Conclusion

C. violaceum appears to be an emerging pathogen whose most human infections are lethal. The scarcity of human infections due to this microorganism, particularly in Africa, contributes to the exacerbation of clinicians' and laboratory staff's lack of awareness of its real impact in case of misdiagnosis and late or inappropriate therapeutic management.

Our findings highlight the need for a 'One Health approach' to monitor and control the expansion of this mesophilic bacterium.

Author Contributions

The conception, study design, execution, acquisition of data, analysis and interpretation (or in all these areas) of the work reported were significantly made by all authors. They have drafted, and substantially revised the article.

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Conflict of Interest

There are no conflicts of interest involving this article.

References

1. Anupop Jitmuang, (2008) Human Chromobacterium violaceum infection in Southeast Asia: case reports and literature review. Southeast Asian J Trop Med Public Health 39(3): 452-460.
2. DC de Lima (2021) Identification of plasmids from Brazilian Chromobacterium violaceum strains. Can J Microbiol pp.1-10.
3. VM de Lima, BB Batista, JF da Silva Neto (2022) The Regulatory Protein ChuP Connects Heme and Siderophore-Mediated Iron Acquisition Systems Required for Chromobacterium violaceum Virulence. Front Cell Infect Microbiol 12: 873536.
4. BPH A Sneath (1956) Cultural and Biochemical Characteristics of the Genus Chromobacterium. J Gen Microbiol 15(1): 70-98.
5. M Enrico, MF Baggi, E Luigia, L Mattia (2020) Chromobacterium violaceum bacteraemia : a new entity in Switzerland. Swiss Med Wkly 150: w20220.

6. E Bottieau (2014) Fatal *Chromobacterium violaceum* bacteraemia in rural Bandundu, Democratic Republic of the Congo. *New Microbes New Infect* 3: 21-23.
7. NP Parajuli (2016) Bacteremia caused by a rare pathogen-*Chromobacterium violaceum*: A case report from Nepal. *Int J Gen Med* 9: 441-446.
8. R Sivendra, HS Lo (1975) Identification of *Chromobacterium violaceum*: Pigmented and Non-pigmented Strains. *J Gen Microbiol* 90(1): 21-31.
9. CC Moore, JE Lane, JL Stephens (2001) Successful treatment of an infant with *Chromobacterium violaceum* sepsis. *Clin Infect Dis* 32(6): E107-E110.
10. D Rettori, N Durán (1998) Production, extraction and purification of violacein: An antibiotic pigment produced by *Chromobacterium violaceum*. *World J Microbiol Biotechnol* vol. 14(5).
11. GZ Justo, N Durán(2017) Action and function of *Chromobacterium violaceum* in health and disease: Violacein as a promising metabolite to counteract gastroenterological diseases. *Best Pract Res Clin Gastroenterol* 31(6): 649-656.
12. KE Aldridge, GT Valainis, CV Sanders (1988) Comparison of the In Vitro Activity of Ciprofloxacin and 24 Other Antimicrobial Agents Against Clinical Strains of *Chromobacterium violaceum*. *Diagn Microbiol Infect Dis* 10(1): 31-39.
13. B Alisjahbana, J Debora, E Susandi, G Darmawan (2021) *Chromobacterium violaceum*: A review of an unexpected scourge. *Int J Gen Med* 14: 3259-3270.
14. CH Yang, YH Li (2011) *Chromobacterium violaceum* infection: A clinical review of an important but neglected infection. *J Chin Med Assoc* 74(10): 435-41.
15. DAC Diaz, DA Martin, NB Ortiz, ALO Monroy, VH Angarita, et al. (2021) *Chromobacterium violaceum* Periareolar Infection, First Non-Lethal Case in Colombia: Case Report and Literature Review. *Infect Dis Rep* 13(2): 571-581.