

**Mini Review**

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Thrombolysis with Tissue Plasminogen Activator (Tpa) Alone has been A Longstanding Mistake

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Current thrombolytic therapy consists of high dose tPA administered iv for 60-90 minutes. This treatment has had only limited success in acute myocardial infarction (AMI) [1-3], and consequently has more recently been replaced by percutaneous coronary intervention (PCI) as the treatment of choice in AMI [4]. Although PCI is time-consuming and delays coronary reperfusion, better clinical outcomes than with tPA were nevertheless obtained [5]. In the treatment of ischemic stroke, a tPA dose reduction was required due to a 20% intracranial hemorrhage (ICH) incidence when the AMI dose was used [6]. Dose reduction diminished the thrombolytic efficacy and resulted in a reperfusion rate of only about 30% [7], and a 7% incidence of symptomatic ICH remained [8]. As in AMI, PCI is now being used increasingly in stroke JTT [9-11]. Historically, the use tPA for thrombolysis was based on the belief that tPA was responsible for intravascular fibrinolysis, whereas the other natural plasminogen activator, urokinase plasminogen activator (uPA), was responsible for extravascular fibrinolysis where its effect is mediated by its uPA receptor, UPAR. Although this belief is rarely questioned, it is belied by a number of findings such as those from gene knockout animal studies. These showed that knocking out the uPA gene, but not the tPA gene, impairs lysis of a venous thrombus. The tPA gene knockout had little effect on thrombus resolution [12]. Similarly, a uPA but not a tPA knockout resulted in spontaneous fibrin formation, and uPA mediated fibrinolysis was not impaired by deletion of the UPAR gene [13], contrary to what had been believed.

The tPA dominance concept was implicitly challenged by these findings and also by the unexpected difficulty that was encountered

to show that tPA was a better thrombolytic than streptokinase (SK), a non-specific, indirect, and inherently flawed plasminogen activator. An unprecedented total of 95,740 patients with AMI, in three mega-trials, were required to show a statistically significant difference [1-3]. Bayesian analysis of these findings suggested that "the clinical superiority of tPA over SK remains uncertain" [14]. Although there is little uPA in plasma, which reinforced the idea that it was an extravascular activator, there is a significant reservoir of uPA on the surface of platelets [15, 16] and monocytes [17], and uPA's half-life is 2.5 days rather 7 minutes. It is also noteworthy that fibrinolysis involves the activation of three different fibrin-bound plasminogens [18], and only one of these is activated by tPA at a fibrin-specific doses [19]. The other two are activated by uPA, one by prouPA and the other by tcuPA [20-22]. Only at high non-specific doses can tPA activate these two plasminogens, and without its fibrin promotion, tPA is a poor plasminogen activator.

These findings help explain the disappointing results obtained with tPA monotherapy. In addition, since tPA is an enzyme, it is also ill-suited for administration by infusion, and due to its high fibrin affinity, this is unnecessary for its therapeutic effect. In biological fibrinolysis, tPA is stored in the vessel wall, and in the event of intravascular thrombus formation, it is released at that site, binds to the fibrin clot, and initiates thrombolysis. Lysis creates two new plasminogen binding sites on fibrin [23]. Plasminogen on one of these is activated specifically by prouPA [20, 24] which is itself activated to tcuPA in the process [21], and tcuPA activates the remaining plasminogen [22]. Therefore, the two activators are complementary, with thrombolysis being initiated by tPA

and completed by uPA [25]. Using tPA alone for thrombolysis is somewhat analogous to trying to run a car on the starting motor. The activators are also synergistic [25] and this was once tested in a clinical study of 101 patients with AMI. Patients were given a mini-bolus of tPA (5 mg) followed by an infusion (40 mg/h) of prouPA. A TMI-3 infarct artery patency of 60% was obtained, with

no reocclusions or ICH and only a 1% mortality [26], results which compare very favorably with the best of the tPA trials in AMI [3, 27]. Therefore, a small bolus of tPA, 5% of the standard dose, was sufficient in the presence of the other activator for exceptionally effective and safe thrombolysis, as also shown experimentally in vitro (Figure 1).

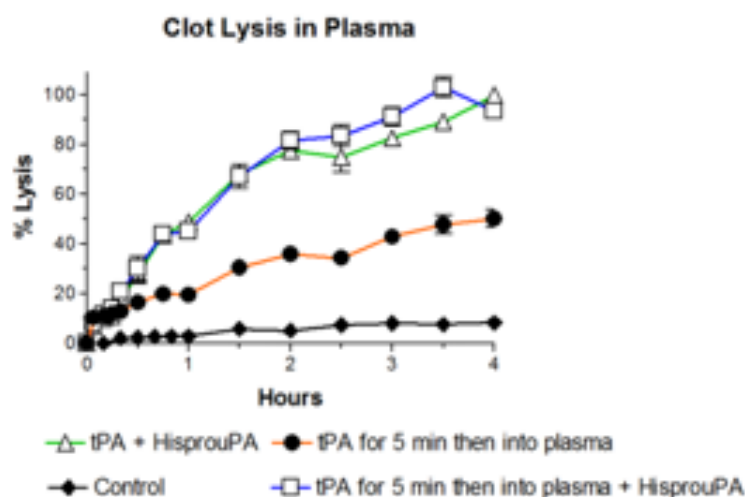


Figure 1: 5 minute exposure of clot to tPA (0.2 µg/ml) followed by wash and then clot put into plasma containing HisprouPA (6 µg/ml) (blue) induces same lysis as when tPA is in plasma throughout lysis (green).

Unfortunately, the above clinical study could not be followed up since prouPA commercial development was abandoned not long thereafter. This was because at the therapeutic doses required when used alone, prouPA became unstable in blood and converted to tcuPA, resulting in bleeding complications. To deal with this problem, a more stable, single site (Lys300→His) mutant prouPA was developed which allows the physiological properties of prouPA to be preserved at therapeutic doses [28-38].

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Conflict of Interest

No conflict of interest.

References

- Gruppo Italiano Per Lo Studio Della Sopravvivenza Nell'Infarto Miocardico (1990) GISSI-2 A factorial randomised trial of alteplase versus streptokinase and heparin versus no heparin among 12,490 patients with acute myocardial infarction. *Lancet* 336: 65-71.
- ISIS-3 Third International Study of Infarct Survival Collaborative Group (1992) ISIS-3: A randomised comparison of streptokinase vs tissue plasminogen activator vs anistreplase and of aspirin plus heparin vs aspirin alone among 41,299 cases of suspected acute myocardial infarction. *The Lancet* 339: 65-71.
- The GUSTO Investigators (1993) An international randomized trial comparing four thrombolytic strategies for acute myocardial infarction. *N Engl J Med* 329: 673-682.
- Keeley EC, Boura JA, Grines CL (2003) Primary angioplasty versus intravenous thrombolytic therapy for acute myocardial infarction: a quantitative review of 23 randomised trials. *Lancet* 361: 13-20.
- Elliott M Antman, Mary Hand, Paul W Armstrong, Eric R Bates, Lee A Green, et al. (2008) 2007 focused update of the ACC/AHA 2004 guidelines for the management of patients with ST-elevation myocardial infarction. *Circulation* 117: 296-329.
- Boysen G and The ECASS Study Group (1995) European cooperative acute stroke study (ECASS): (rt-PA - thrombolysis in acute stroke) study design and progress report. *Eur J Neurol* 1: 213-219.
- The National Institute of Neurological Disorders and Stroke rt-PA Stroke Study Group (1995) Tissue plasminogen activator for acute ischemic stroke. *N Engl J Med* 333: 1581-1587.
- (2012) The benefits and harms of intravenous thrombolysis with recombinant tissue plasminogen activator within 6 h of acute ischemic stroke (the third international stroke trial [IST-3]: a randomized controlled trial *The Lancet* 379(9834): 2352-2363.
- Bruce CV Campbell, Peter J Mitchell, Timothy J Kleinig, Helen M Dewey, Leonid Churilov, et al. (2015) Endovascular therapy for ischemic stroke with perfusion-imaging selection. *N Engl J Med* 372: 1009-1018.
- Berkhemer OA, Fransen PSS, Beumer D, Lucie A van den Berg, Hester F Lingsma, et al. (2015) A randomized trial of intraarterial treatment for acute ischemic stroke. *N Engl J Med* 372: 11-20.
- Goyal M, Demchuk AM, Menon BK, Muneer Eesa, Jeremy L Rempel, et al. (2015) Randomized assessment of rapid endovascular treatment of ischemic stroke. *N Engl J Med* 372: 1019-1030.
- Singh I, Burnand KG, Collins M, Luttun A, Collen D, et al. (2003) Failure of thrombus to resolve in urokinase-type plasminogen activator gene-knockout mice: rescue by normal bone marrow-derived cells. *Circulation* 107: 869-875.
- Bugge TH, Flick MJ, Danton MJ, Daugherty CC, Rømer J, et al. (1996) Urokinase-type plasminogen activator is effective in fibrin clearance in the absence of its receptor or tissue-type plasminogen activator. *Proc Natl Acad Sci USA* 93: 5899-5904.
- Brophy JM, Joseph L (1995) Placing trials in context; GUSTO visited by Reverend Bayes. *JAMA* 273: 871-875.

15. Vaughan DE, Vautloutle E, Collen D (1990) Urokinase binds to platelets through a specific saturable, low affinity mechanism. *Fibrinolysis* 4: 141-146.
16. Gurewich V, Johnstone M, Loza JP, Pannell R (1993) Pro-urokinase and prekallikrein are both associated with platelets: implications for the intrinsic pathway of fibrinolysis and for therapeutic thrombolysis. *FEBS Lett* 318: 317-321.
17. Grau E, Moroz LA (1989) Fibrinolytic activity of normal human blood monocytes. *Thromb Res* 53: 145-162.
18. Suenson E, Lützen O, Thorsen S (1984) Initial plasmin-degradation of fibrin as the basis of a positive feed-back mechanism in fibrinolysis. *Eur J Biochem* 140: 513-522.
19. Hoylaerts M, Rijken DC, Lijnen HR, Collen D (1982) Kinetics of the activation of plasminogen by human tissue plasminogen activator. Role of fibrin. *J Biol Chem* 257: 2912-2929.
20. Liu J, Gurewich V (1992) Fragment E-2 from fibrin substantially enhances pro-urokinase-induced glu-plasminogen activation. A kinetic study using a plasmin-resistant mutant pro-urokinase (A-158-rpro-UK). *Biochemistry* 31: 6311-6317.
21. Petersen LC (1997) Kinetics of reciprocal pro-urokinase/plasminogen activation. Stimulation by a template formed by the urokinase receptor bound to poly (D-lysine). *Eur J Biochem* 245: 316-323.
22. Gurewich V (2015) Why so little progress in therapeutic thrombolysis? The current state of the art and prospects for improvement. *J Thromb Thrombolysis* 40: 480-487.
23. Harpel PC, Chang TS, Verderber E (1985) Tissue plasminogen activator and urokinase mediate the binding of Glu-plasminogen to plasma fibrin I. Evidence for new binding site in plasmin-degraded fibrin I. *J Biol Chem* 260: 4432-4440.
24. Liu J, Gurewich V (1991) A comparative study of the promotion of tissue plasminogen activator and pro-urokinase-induced plasminogen activation by fragments D and E-2 of fibrin. *J Clin Invest* 88: 2012-2017.
25. Pannell R, Black J, Gurewich V (1988) The complementary modes of action of tissue plasminogen activator (t-PA) and pro-urokinase (pro-UK) by which their synergistic effect on clot lysis may be explained. *J Clin Invest* 81: 853-859.
26. Zarich SW, Kowalchuk GJ, Weaver WD, Loscalzo J, Sassower M, et al. (1995) Sequential combination of thrombolytic therapy for acute myocardial infarction: results of the pro-urokinase and t-PA enhancement of thrombolysis (PATENT) trial. *J Am Coll Cardiol* 26: 374-379.
27. Ross AM for the GUSTO Angiographic Investigators (1993) The effects of tissue plasminogen activator, streptokinase, or both on coronary-artery patency, ventricular function, and survival after acute myocardial infarction. *N Engl J Med* 329: 1615-1622.
28. Liu JN, Tang W, Sun ZY, Kung W, Pannell R, et al. (1996) A site-directed mutagenesis of pro-urokinase which substantially reduces its intrinsic activity. *Biochemistry* 35: 14070-14076.
29. Sun Z, Jiang Y, Ma Z, Wu H, Liu BF, et al. (1997) Identification of a flexible loop (297-313) of urokinase-type plasminogen activator, which helps determine its catalytic activity. *J Biol Chem* 272: 23818-23823.
30. Tang W, Sun Z, Pannell R, Gurewich V, Liu J (1997) An efficient system for production of recombinant urokinase-type plasminogen activator. *Protein Expr Purif* 11: 279-283.
31. Sun Z, Liu BF, Chen Y, Gurewich V, Zhu D, et al. (1998) Analysis of the forces which stabilizes the active conformation of urokinase-type plasminogen activator. *Biochemistry* 37: 2935-2940.
32. Liu JN, Liu JX, Liu B, Sun Z, Zuo JL, et al. (2002) A prourokinase mutant which induces highly effective clot lysis without interfering with hemostasis. *Circu Res* 90: 757-763.
33. Gurewich V, Pannell R, Simmons-Byrd A, Sarmientos P, Liu JN, et al. (2006) Thrombolysis versus bleeding from hemostatic sites by a prourokinase mutant compared with tissue plasminogen activator. *J Thrombos Haemostas* 4: 1559-1565.
34. Pannell R, Kung W, Gurewich V (2007) C1-inhibitor prevents non-specific plasminogen activation by a prourokinase mutant without impeding fibrin-specific fibrinolysis. *J Thrombos Haemostas* 5: 1047-1054.
35. Pannell R, Kung W, Gurewich V (2007) C1-inhibitor prevents non-specific plasminogen activation by a prourokinase mutant without impeding fibrin-specific fibrinolysis. *J Thrombos Haemostas* 5: 1047-1054.
36. Gurewich V, Pannell R (2009) Recombinant human C1-inhibitor prevents non-specific proteolysis by mutant prouPA during optimal fibrinolysis. *Thrombos Haemostas* 102(2): 279-286.
37. Tomasi S, Sarmientos P, Giorda G, Gurewich V, Vercelli A (2011) Mutant prourokinase with adjunctive C1-inhibitor is an effective and safer alternative to tPA in rat stroke. *PLoS One* 6: e21999.
38. Pannell R, Li S, Gurewich V (2015) Highly effective fibrinolysis by a sequential synergistic combination of mini-dose tPA plus low-dose mutant proUK. *PLOS ONE* 10(3): e122018.